

Article

Co-Application of 24-Epibrassinolide and Titanium Oxide Nanoparticles Promotes *Pleioblastus pygmaeus* Plant Tolerance to Cu and Cd Toxicity by Increasing Antioxidant Activity and Photosynthetic Capacity and Reducing Heavy Metal Accumulation and Translocation

Abolghassem Emamverdian ^{1,2,*}, Yulong Ding ^{1,2}, James Barker ³, Guohua Liu ^{1,2,*}, Mirza Hasanuzzaman ^{4,*}, Yang Li ⁵, Muthusamy Ramakrishnan ^{1,2} and Farzad Mokhberdoran ¹

- ¹ Co-Innovation Center for Sustainable Forestry in Southern China, Nanjing Forestry University, Nanjing 210037, China; ylding@njfu.com.cn (Y.D.); ramky@njfu.edu.cn (M.R.); mfarzad649@hotmail.com (F.M.)
- ² Bamboo Research Institute, Nanjing Forestry University, Nanjing 210037, China
- ³ School of Life Sciences, Pharmacy and Chemistry, Kingston University, Kingston-upon-Thames, Surrey KT1 2EE, UK; j.barker@kingston.ac.uk
- Department of Agronomy, Faculty of Agriculture, Sher-e-Bangla Agricultural University, Dhaka 1207, Bangladesh
- Department of Mathematical Sciences, Florida Atlantic University, Boca Raton, FL 33431, USA; yangli@fau.edu
- Correspondence: emamverdiyan@njfu.edu.cn (A.E.); ghliu@njfu.edu.cn (G.L.); mhzsauag@yahoo.com (M.H.); Tel.: +86-1585-0741-241 (A.E.)

Abstract: The integrated application of nanoparticles and phytohormones was explored in this study as a potentially eco-friendly remediation strategy to mitigate heavy metal toxicity in a bamboo species (*Pleioblastus pygmaeus*) by utilizing titanium oxide nanoparticles (TiO₂-NPs) and 24-epibrassinolide (EBL). Hence, an in vitro experiment was performed to evaluate the role of 100 μ M TiO₂ NPs and 10^{-8} M 24-epibrassinolide individually and in combination under 100 μ M Cu and Cd in a completely randomized design using four replicates. Whereas 100 μ M of Cu and Cd reduced antioxidant activity, photosynthetic capacity, plant tolerance, and ultimately plant growth, the co-application of 100 μ M TiO₂ NPs and 10^{-8} M EBL+ heavy metals (Cu and Cd) resulted in a significant increase in plant antioxidant activity (85%), nonenzymatic antioxidant activities (47%), photosynthetic pigments (43%), fluorescence parameters (68%), plant growth (39%), and plant tolerance (41%) and a significant reduction in the contents of malondialdehyde (45%), hydrogen peroxide (36%), superoxide radical (62%), and soluble protein (28%), as well as the percentage of electrolyte leakage (49%), relative to the control. Moreover, heavy metal accumulation and translocation were reduced by TiO₂ NPs and EBL individually and in combination, which could improve bamboo plant tolerance.

Keywords: toxic metals/metalloid; nanoparticles; phytohormones; phytoremediation; reactive oxygen species

1. Introduction

In recent decades, increasing anthropogenic activities have led to increases in greenhouse gases in the natural environment, and chemical fertilization has led to increases in heavy metal contamination in forestland and agricultural soils, deleteriously contributing to global climate change [1]. Many reports have shown that heavy metals pose a major threat to agricultural land, animals, and plants, which can influence the human food chain, leading to negative effects on human health [2,3]. Copper (Cu) and cadmium (Cd) have been mentioned as being the most abundant toxic metals in Chinese farmland soils [4].



Citation: Emamverdian, A.; Ding, Y.; Barker, J.; Liu, G.; Hasanuzzaman, M.; Li, Y.; Ramakrishnan, M.; Mokhberdoran, F. Co-Application of 24-Epibrassinolide and Titanium Oxide Nanoparticles Promotes *Pleioblastus pygmaeus* Plant Tolerance to Cu and Cd Toxicity by Increasing Antioxidant Activity and Photosynthetic Capacity and Reducing Heavy Metal Accumulation and Translocation. *Antioxidants* 2022, *11*, 451. https:// doi.org/10.3390/antiox11030451

Academic Editor: Nafees A. Khan

Received: 25 January 2022 Accepted: 20 February 2022 Published: 24 February 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). While there is no evidence of the biological activity of Cd in the plant growth process, trace amounts of Cu could have a positive impact as a dietary nutrient on plant growth; however, extreme Cu levels induce plant toxicity [5]. Cu, as a trace element, can enhance photosynthetic efficiency, such as electron transport. Additionally, Cu regulates structural proteins involved in cell wall metabolism and can elevate mitochondrial respiration to produce energy [6]. Conversely, surplus concentrations of Cu in the form of Cu²⁺ are responsible for oxidative stress due to the generation of reactive oxygen species (ROS) compounds [7], which can result in a reduction in plant growth with an altered functionality of the cell membrane, limitation of enzyme activities, and depression of photosynthetic efficiency, ultimately leading to plant death [8]. Cd, a nonessential element with high toxicity, is known as the most dispersed element in soils and irrigation water, which has a destructive influence on plant and human life [9]. Excess Cd in soil and the absorption of cadmium by plants increase ROS production, such as free radicals, which are the main factors in the initiation of oxidative stress in plants [9]. Cadmium has a damaging impact on plant cell functions and the metabolic pathways involved in the production of lipids, proteins, and nucleic acids. Cd injures the cell membrane, which leads to lipoperoxidation and oxidative toxicity in plants. It has been demonstrated that cadmium reduces the plant defense system with a reduction in antioxidant activity capacity. This phenomenon finally reduces plant photosynthesis and inhibits plant growth and development [9].

Nanoparticles with unique structures and sizes (1 to 100 nm) [10] have been observed to increase plant nutrients and crop production [11]. The diverse surface-to-volume ratios of nanoparticles could differ from their bulk counterparts [12]. Recently, some studies have reported that titanium nanoparticles (TiO_2) have the ability to increase plant growth under metal stress [13–17]. Therefore, we suggest that titanium could be a good material to reduce plant stress. Brassinosteroids (BRs) are a new phytohormone and belong to the polyhydroxy steroidal group. There are 70 types of BRs in plants. Among them, 24-epibrassinolide (EBL) is known as the top bioactive BR that can promote plant growth under stressful conditions [18]. In addition, 24-epibrassinolide EBL induces antioxidant activity, plant photosynthesis, seed yield, and oxidative production under stressful conditions [19–21]. It has been indicated that the interaction of EBL with other cellular molecules can enhance signaling efficiency within the plant defense grid under stress conditions [22]. This phenomenon can boost antioxidant capacity in the face of multiple stressful factors, such as HMs [23]. This research study represents the individual and co-application of TiO_2 -NPs and EBL, as well as the investigation of their role in the alleviation of Cu and Cd toxicity in bamboo plants with an emphasis on antioxidant, photosynthetic, and plant growth parameters.

Bamboo (Bambusoideae) species occupy the largest portion of Chinese farmland (6 million hectares) [24,25]. This fast-growing plant provides nutrient sources for local family livelihoods in southern and western China [26]. *Pleioblastus pygmaeus* is a suitable species for landscape purposes, with a characteristic height of 30–50 cm. *Pleioblastus pygmaeus* originated in Japan but was transferred to China in the early 20th century. A desirable condition of this plant for this experiment was its adaptation to basic (alkaline), acidic, and neutral soils [27]. Conversely, the excess of heavy metals (frequently Cu and Cd) caused by anthropogenic activities has become a major dilemma for agricultural and forestry soils in this area [4], which can influence bamboo plant growth and development. Hence, it is essential to find appropriate biologic materials to reduce soil toxicity and increase plant tolerance under heavy metal toxicity. Therefore, we selected two applications of TiO_2 NPs and 24-epibrassinolide, individually and in combination, against heavy metal toxicity, which could aid in understanding the involved mechanisms in the combined application of nanoparticles and phytohormones against heavy metal toxicity. To our knowledge, this is the first comprehensive study to investigate the combination of TiO_2 NPs and EBL in the amelioration of Cu and Cd toxicity in bamboo species. Therefore, in this paper, we aim to investigate the impact of TiO_2 NPs and EBL on enhancing plant tolerance under heavy metal toxicity with an emphasis on antioxidant and nonantioxidant enzyme capacity, ROS production, photosynthesis, and growth indices under Cu and Cd.

2. Materials and Methods

2.1. Plant Material and In Vitro Conditions

This research study was performed under in vitro conditions in a plant tissue culture laboratory using MS medium (Murashige and Skoog, 1962) [28] consisting of 6benzylaminopurine (6-BA) (4 mL), micronutrients (10 mL), macronutrients (100 mL), kinetin (KT) (0.5 mL), sucrose (30 g) and agar (8 g) at pH 5.8 \pm 0.1. For this purpose, a completely randomized design (CRD) was employed that contained 100 μ M TiO₂ NPs and 10⁻⁸ M 24 epibrassinolide individually and in combination with 100 μ M Cu as well as 100 μ M Cd in four replications (Table 1). We adjusted the pH value in MS to 5.8 for two reasons: firstly, to optimize nutrient absorption, the availability of the nutrients to the plants was optimum at pH 5.8; and secondly, the preparation of the gelling of the agar-solidified medium should be completed at ca. pH 5.8. To proliferate bamboo roots, young shoots (10 mm long nodal explants) were planted in MS medium supplemented with pyridoxine (3 µM), nicotinic acid (4 μ M), thiamine–HCl (1.2 μ M), myo-inositol (0.6 mM), 30 g L⁻¹ sucrose, and 0.1 mg L⁻¹ indole-3-acetic acid (IAA) as a regulator hormone involved in plant growth. The appropriate amount of each treatment (100 μ M TiO₂ NPs and 10⁻⁸ M 24-epibrassinolide) was mixed in 1 L MS medium, adjusted to pH 5.8 \pm 0.1, and then applied to 8–10 g/L agar. The solution was placed in 60 mm diameter glass petri dishes containing 100 mL of culture, and sterilization of the intended MS medium was conducted in an autoclave (HiClave HVE-50, ZEALWAY-USA, Delaware, DE, USA) at the optimum temperature of 110 °C for 40 min. The dishes were transferred to an Air Tech incubation hood with ultraviolet sterilization with white fluorescent lamps (wavelength between 10 and 420 nm) at a temperature of 25 °C for 4 h. In the final step, the plantlet treatments were preserved as research materials in a controlled tissue culture chamber with fluorescent lamps (white) at a wavelength between 10 and 420 nm. In terms of temperature, the growth was performed at 17/22 °C in the dark periods and 30/25 °C in the light periods for three weeks.

Treatments	Concentrations		
Control	0		
Cu	100 μM Cu		
Cd	100 µM Cd		
TiO ₂	100 μM TiO ₂		
$TiO_2 + Cu$	100 μM TiO ₂ + 100 μM Cu		
$TiO_2 + Cd$	100 μM TiO ₂ + 100 μM Cd		
EBL	10^{-8} M EBL		
EBL + Cu	10^{-8} M EBL + 100 μ M Cu		
EBL + Cd	10^{-8} M EBL + 100 μ M Cd		
$TiO_2 + EBL$	$100 \ \mu M TiO_2 + 10^{-8} M EBL$		
$TiO_2 + EBL + Cu$	$100 \ \mu M TiO_2 + 10^{-8} M EBL + 100 \ \mu M Cu$		
$TiO_2 + EBL + Cd$	$100 \ \mu M \ TiO_2 + 10^{-8} \ M \ EBL + 100 \ \mu M \ Cd$		

Table 1. The treatment combinations of the experiment.

Titanium nanoparticles were provided by Nanjing Jiancheng Company, Jiangsu Province, China, and consisted of a white powder with a purity of >99% nanotitanium and a diameter of 25 nm. The levels of Cu and Cd were chosen according to the previous studies, which displayed high and low levels of toxicity in bamboo plants [13,14]. Bamboo (*A. pygmaeus*) was selected from local species by the Bamboo Research Institute, which is located at Nanjing Forestry University.

In this research study, biomass and growth indices, including root and shoot dry weight (DW) and shoot length, were quantified. To investigate photosynthesis pigments, total chlorophyll (Chl), Chl a and b, and carotenoid contents were measured. To determine the fluorescence parameters, 5 parameters were recorded, including: (i) actual photochemical efficiency of PSII (ϕ PSII), (ii) maximum photochemical efficiency of PSII (Fv/Fm), (iii) photochemical quenching coefficient (qP), (iv) effective photochemical efficiency of

PSII (Fv'/Fm'), and (v) nonphotochemical quenching (NPQ). Heavy metal accumulation and TiO_2 NP contents were measured in leaves, stems, and roots. Plant defense enzymes and nonenzymatic antioxidants were measured. To assay cell membrane injury, ROS compounds, electron leakage, and malondialdehyde (MDA) content were estimated. Finally, the translocation factor (TF), bioaccumulation factor (BAF), and tolerance index (TI) of the shoots and roots were calculated.

2.2. Preparation of Samples

Leaf samples were collected from the different treatments, and then 0.5 g samples were placed in a container and crushed into a powder. An appropriate amount of liquid nitrogen was added to the samples, and the obtained powder was dissolved in PBS (pH 7.2–7.4) at 2–8 °C. The solution was centrifuged at $2500-3500 \times g$ for 17 min to extract the supernatant, which was kept for use in antioxidant enzyme activity tests.

2.3. Protective Enzymes

Superoxide dismutase (SOD, EC: 1.15.1.1) was measured based on the results of photoreduction obtained by nitro blue tetrazolium (NBT), which was conducted using the Zhang method [29]. Peroxidase (POX, EC: 1.11.1.7) was estimated by using the protocol of Upadhyaya [30]. Catalase (CAT, EC: 1.11.1.6) was estimated based on the results of reactions analyzing H_2O_2 at an absorbance of 240 nm, which was estimated by the Aebi protocol [31]. Glutathione reductase (GR, EC: 1.6.4.2) was estimated using the protocol reported by Foyer and Halliwell [32] with some modifications. Ascorbate peroxidase (APX, EC: 1.11.1.1) was measured using the Nakano and Asada method [33]. APX antioxidant activity was obtained by recording the reduction in absorbance at 290 nm (coefficient of absorbance at 2.8 mM⁻¹ cm⁻¹). Phenylalanine ammonia-lyase (PAL, EC: 4.3.1.5) activity was assessed using the Berner [34] protocol.

2.4. Assessment of Nonenzymatic Antioxidant Activities (Flavonols, Tocopherols, and Total Phenolics)

Methanolic Extract Preparation

For this test, 0.5 g of dry leaf sample was dissolved in 4 mL of methanol (80%) and then centrifuged at 7000 \times g for 15 min. The methanolic extract was used for the tests. The total phenolics were measured according to the protocol of Conde [35]. According to this protocol, a 0.1-m methanolic extra was added to 2.5 mL of 10% Folin–Ciocalteu reagent. Then, for neutralization of the obtained mixture of sodium bicarbonate, 7% was added. The final mixture was transferred to a spectrometer machine to measure the total phenolics at an absorbance of 765 nm. The content of flavonol was determined according to the Akkol method [36]. A 0.5-mL methanolic extract was homogenized with 0.4 mL of aluminum chloride (2%) and 1.5 mL of sodium acetate (5%). After preparation of the supernatant, it was kept at room temperature for 2.5 h. The flavonoid content was determined in the supernatant at an absorbance of 445 nm. The content of tocopherol was determined according to the protocol of Kayden [37]. For this purpose, 3 mL ethanol was mixed with 0.1 g of leaf samples, and the soluble solution was then centrifuged at $7000 \times g$ for 15 min. The obtained mixture was added to 0.1 mL ethanol extract, 0.2 mL bathophenanthroline at a concentration of 0.2%, 0.001 M of 0.2 mL ferric chloride, and 1 mM 0.2 mL phosphoric acid. The content of tocopherol was recorded by measuring the absorbance of the supernatant at 534 nm.

2.5. Assay of Hydrogen Peroxide (H_2O_2), Malondialdehyde (MDA), Superoxide Radical ($O_2^{\bullet-}$), Soluble Proteins (SP), and Electrolyte Leakage (EL)

Malondialdehyde is representative of lipid peroxidation, which was measured by the protocol described by Siddiqui [38]. In this experiment, 0.1% trichloroacetic acid (TCA) was used for the homogenization of fresh leaves, after which the sample was centrifuged at $8000 \times g$ for 25 min. The obtained amount of supernatant was mixed with TCA solution in the range of 20%, which contained 0.5% thiobarbituric acid. In the next process, the

soluble solution was kept at 98 °C for 25 min. Then, the soluble solution was kept at room temperature. The final soluble solution was centrifuged a second time at $2000 \times g$ for 15 min at 5 °C. Finally, to estimate malondialdehyde, the absorbance was determined at 532 nm.

The levels of H_2O_2 were determined using the protocol reported by Patterson [39]. For this study, samples (leaves) in the specified amount of 0.5 g were mixed in a mortar and pestle by adding 10 mL cold acetone. The mixture was centrifuged at $4000 \times g$ for 25 min. In the next step, titanium chloride at a concentration of 20% in 2 mL of concentrated HCl and 2 mL of ammonia at the specified level of 17 M were added to the supernatant (1 mL). The supernatant was extracted with acetone, which was conducted by the addition of 2 N H₂SO₄ in 10 mL for proper absorbance. To remove immiscible inputs, the mixture was centrifuged again. The absorbance of the supernatant was recorded at 410 nm. The levels of H_2O_2 were determined based on a standard curve, which was created based on the known levels of H_2O_2 and formulated as µmole g^{-1} FM. The soluble protein (SP) levels were assigned according to the protocol of Bradford [40] and measured based on the effect of Coomassie Brilliant Blue (G25) on changes in protein levels. The final data were obtained using a spectrometer machine. The amount of superoxide radical $(O_2^{\bullet-})$ was determined according to the method of Li [41]. According to this protocol, 200 mg leaf tissue samples were mixed with phosphate buffer at pH 7.8 in the amount of 65 mM and then centrifuged at 4000 \times g for 20 min. The supernatant was incubated in 10 mM of hydroxylamine hydrochloride and 65 mM of phosphate buffer (pH = 7.8) for 15 min at 27 °C. In the next step, 7 mM α -naphthylamine plus 17 mM sulfanilamide was added to the mixture, preserved for 25 min and then recorded at an absorbance of 530 nm at 25 $^\circ$ C. Finally, to determine the final rate of $O_2^{\bullet-}$, nitrogen dioxide radicals (NO₂) were applied to generate a standard curve. Electrolyte leakage (EL) was calculated based on the protocol of Valentovic [42]. According to this protocol, 0.3 g of leaf samples were mixed with 15 mL of deionized water. Then, the mixture was kept at the optimum temperature (25 $^{\circ}$ C) for 2.5 h. In this stage, EC_1 was recorded as the primary electrical conductivity of the mixture. To obtain EC_2 as the secondary electrical conductivity, the samples were transferred to one autoclave and kept at 120 °C for 17 min. At the end of the test, EL was determined based on the following formula:

$$EL(\%) = EC_1 / EC_2 \times 100 \tag{1}$$

2.6. Measurement of Photosynthetic Pigments and Fluorescence Parameters

Photosynthetic pigments, such as Chl a and b, and carotenoid levels were determined according to the protocol of Lichtenthaler and Buschmann [43]. For pre-experiment bamboo samples, ca. 0.5 g was provided, and then the samples were transferred to a mortar with liquid nitrogen. To prepare the liquid sample extract, the obtained powder was mixed with 20 mL of acetone at a specific concentration of 80% at 0 to 5 °C. Then, it was centrifuged at $5000 \times g$ for 15 min. At the end of the experiment, Chl a, Chl b, and carotenoid levels were determined at absorbances of 663, 645, and 470 nm, respectively. Finally, the levels of Chl and carotenoids were calculated based on the following formulae, which were set in units equal to mg/g fresh weight:

Chlorophyll a =
$$12.25A663 - 2.79A647$$
 (2)

Chlorophyll b =
$$21.50A647 - 5.10A663$$
 (3)

$$Total Chl = Chl a + Chl b$$
(4)

Carotenoid =
$$1000A470 - 1.82Chl a - 95.15 Chl b/225$$
 (5)

A chlorophyll fluorescence imager (CFI) (England) was used to measure fluorescence characteristics, which was conducted under specific dark-adapted conditions for 35 min. To measure the light fluorescence parameters, the fluorescence characteristics were recorded at 700 micromoles $m^{-2} s^{-0}$ in an illumination incubator for actinic light activation. In this study, the main fluorescence indices were: (i) actual photochemical efficiency of

PSll (φ PSll); (ii) effective photochemical efficiency of PSll (Fv'/Fm'); (iii) photochemical quenching coefficient (qP); (4) maximum photochemical efficiency of PSll (Fv/Fm); and (5) non-photochemical quenching (NPQ).

2.7. Measurement of TiO₂ NPs and Metal Accumulation in Roots, Stems, and Leaves of Bamboo

Copper (Cu) and cadmium (Cd) contents and titanium accumulation in roots, stems, and leaves of bamboo species were investigated. For their measurement, the samples were cleaned and dried in an oven, and subsequently 70% nitric acid was added to the samples, which were preserved at an optimum temperature of 70 °C for 20 min. Then, the obtained solution was centrifuged at $9000 \times g$ for 20 min. The contents of Cu and Cd in the plant organs, such as roots, stems, and leaves, were assigned and analyzed using an atomic absorption spectrometry machine (AAS HITACHI, High-Tech Company Tokyo, Japan). In this process, a spectrometer equipped with a furnace of graphite and correction system of the Zeeman-effect background was performed (AAnalyst 800, Perkin Elmer, Norwalk, CT, USA). For determination of the element contents, the different characteristics of the instruments were adjusted. Standardization of the metal was performed using 2.5% nitric acid (spectra scan). For calibration, confirmation of the standard (Perkin Elmer), which contained all of the elements in one inorganic target analyst list (TAL), was performed at optimum intervals in one unattended automatic analysis run.

2.8. Biomass Determination and Shoot Length

Plant biomass was determined by measuring the dry weight of roots and shoots. Firstly, samples (root and shoot treatment) were cleaned and then placed in an oven to remove the water from their surfaces during the process. The samples were fixed at $115 \degree$ C for 25 min. To determine the plant dry weight, the samples were maintained at 70 \degree C for 10 h and then weighed. To determine the shoot length, bamboo plants were measured at the beginning and end of the study. The final data were defined as the difference in plant height.

2.9. Determination of the Tolerance Index (TI), Bioaccumulation Factor (BAF), and Translocation Factor (TF)

To determine the plant tolerance to Cu and Cd toxicity, the indices of the tolerance index (TI), bioaccumulation factor (BAF), and translocation factor (TF) were calculated. This result was obtained based on the method of Souri and Karimi [44] and is considered to denote the efficiency of photoextraction. The formula below is used to find the value of TI, which represents the tolerance index in the shoot and root; TF, which represents the translocation factor; and BAF, which represents the bioaccumulation factor of leaves, stem, and root.

TF (leaves/stem) = the concentration of the co-application of Ti-EBL/heavy	
metals (Cu, Cd) in the leaves/stem of plants (mg/kg)/the concentration of	(6)
the co-application of Ti-EBL/heavy metals (Cu, Cd) in the roots of plants	

TI shoot/root = the dry weight of plant shoot/root from the co-application of Ti-EBL/heavy metal (Cu, Cd) treatment (g)/the dry weight of plant (7) shoot/root from the control (g)

BF (leaves/stem/root) = concentrations of heavy metals in the leaves or stem or root/concentrations of heavy metals in the medium (8)

2.10. Statistical Analysis

A completely randomized design (CRD) was used in this study, consisting of a 2-way factorial with four replicates. R software was used for ANOVA (analysis of variance), and the Tukey's test was used to determine mean differences, which were conducted at the p < 0.05 probability level.

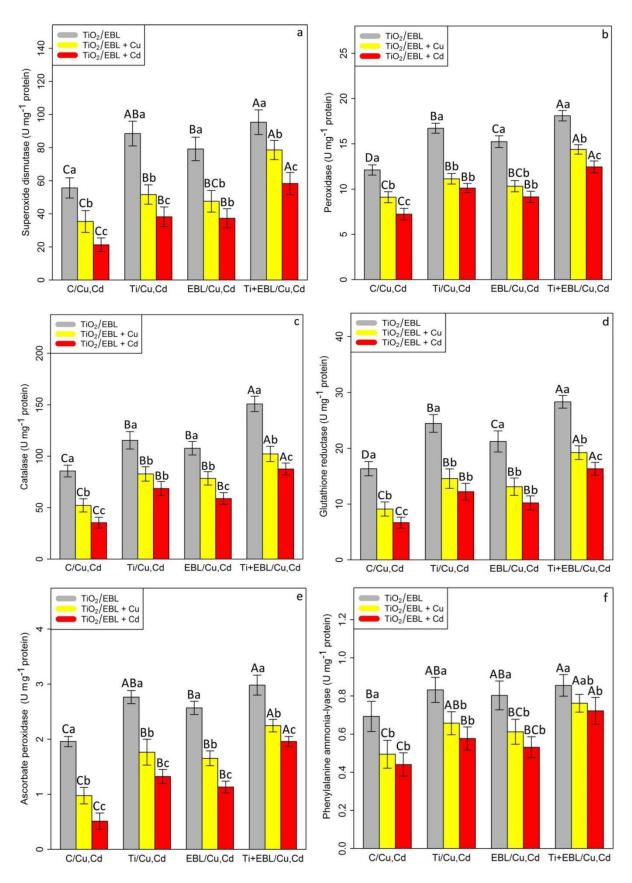
3. Results

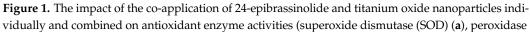
3.1. 24-Epibrasinolide and Titanium Oxide Nanoparticles Promote Antioxidant Capacity in Plants under Cu and Cd Toxicity

The data analysis revealed a significant difference between the various treatments of the co-application of EBL–TiO₂ NPs with Cu and Cd (p < 0.001). According to the obtained data, TiO₂ and EBL individually could increase antioxidant activity under stress conditions. However, the greatest stimulation of antioxidant activity was observed with the combination of TiO₂ and EBL. The combination of TiO₂ and EBL showed the highest capacity for antioxidant activity stimulation, with a 1.71-fold enhancement of SOD, 1.49-fold increase in POX, 1.76-fold increase in CAT, 1.52-fold increase in APX, 1.73-fold enhancement of GR, and 1.23-fold enhancement of PAL activity in comparison with their control treatments (Figure 1). Conversely, the lowest amount of antioxidant activity was observed with 100 μ M Cu and 100 μ M Cd, which resulted in 36% and 61% reductions in SOD, 24% and 40% reductions in POX, 39% and 58% reductions in CAT, 50% and 73% reductions in APX, 44% and 59% reductions in GR, and 28% and 36% reductions in PAL activities, respectively, compared with the control treatment. According to these results, we suggest that TiO₂ and EBL individually have the potential to reduce Cu and Cd toxicity, but the combination of TiO₂ and EBL has a larger impact on the amelioration of heavy metal toxicity.

3.2. 24-Epibrasinolide and Titanium Oxide Nanoparticles Reduce Malondialdehyde (MDA), Soluble Proteins (SP), Electrolyte Leakage (EL), Hydrogen Peroxide (H_2O_2), and Superoxide Radicals ($O_2^{\bullet-}$)

Our results showed that the TiO₂ NP and EBL concentrations had the ability to reduce ROS compounds and prevent cell membrane injury. The data analyses showed a significant difference in the co-application of TiO₂ NPs and EBL based on the concentration of Cu and Cd in the indices of MDA, H₂O₂, SP, EL, and O₂^{•-} (p < 0.001). The most positive effect of the treatments on heavy metals was related to the combination of TiO₂–EBL with Cu and TiO₂–EBL with Cd, which demonstrated 49% and 42% reductions in MDA content, 38% and 33% reductions in H₂O₂ content, 38% and 36% reductions in O₂^{•-} content, 26% and 25% reductions in SP content, and 50% and 44% reductions in EL, respectively. Additionally, the levels of TiO₂ and EBL individually showed a positive role in the amelioration of oxidative stress, which occurred by restraining ROS production, which in turn resulted in protecting the cell membrane against oxidative free radicals. Conversely, the results showed that 100 µM Cu and 100 µM Cd increased the levels of MDA, H₂O₂, O₂^{•-}, SP, and EL, with 48% and 60% increases in MDA content, 35% and 49% increases in H₂O₂, 28% and 41% increases in O₂^{•-}, 25% and 29% increases in SP, and 50% and 63% increases in EL, respectively, compared with their control treatments (Figure 2).





(POX) (b), catalase (CAT) (c), glutathione reductase (GR) (d), ascorbate peroxidase (APX) (e), and phenylalanine ammonia-lyase (PAL) (f)) in bamboo species (*Pleioblastus pygmaeus*) with 100 μ M Cu and 100 µM Cd. In this study, 1-year-old branches of P. pygmaeus were used as plant treatments together with 100 μ M TiO₂ NPs and 10⁻⁸ M 24-epibrassinolide, individually and in combination with 100 μ M Cu and 100 μ M Cd using four replications. Planting of the treated bamboo was performed in an Air Tech inoculation hood with fluorescent white lamps and ultraviolet light (wavelengths of 10-400 nm) at 15 °C and 30 °C. The bamboo plants were constantly exposed to excess heavy metals for three weeks. Sampling for the measurement of antioxidant enzyme activity (a-f) was conducted after three weeks of plant exposure to the co-application of 24-epibrassinolide and titanium oxide nanoparticles under 100 μ M Cu and 100 μ M Cd. The capital letters (A-C) indicate significant differences between treatments of control (C), titanium (Ti), 24-epibrassinolide (EBL), and 24-epibrassinolide involving individual or combined application of titanium oxide nanoparticles (EBL-TiO₂ NPs) under 100 µM Cu and 100 µM Cd (the bars with similar colors), while the lowercase letters (a-c) denote statistically significant differences at each concentration of the co-application of EBL and TiO₂ NPs, individually or in combination with 100 μ M Cu and 100 μ M Cd (the bars with various colors) based on Tukey's test (p < 0.05).

3.3. 24-Epibrasinolide and Titanium Oxide Nanoparticles Increase Nonenzymatic Antioxidant Activities (Flavonol, Tocopherol, and Total Phenolics) in Bamboo Species under Cu and Cd Toxicity

The effects of TiO₂ NPs and EBL concentrations on nonenzymatic activity (flavonol, tocopherol, and total phenolics) in the bamboo species revealed a significant difference between the co-application of 24-epibrassinolide and titanium oxide nanoparticles with Cu and Cd (p < 0.001). According to the results, the combination of TiO₂-HMs and EBL-HMs significantly increased nonenzymatic antioxidant activities in our bamboo species. However, the greatest increase in nonenzymatic activity under heavy metal stress was related to the combination of TiO₂ –EBL with Cu and TiO₂–EBL with Cd, with a 1.55-fold and 1.51-fold enhancement in flavonols, 1.53-fold and 1.51-fold enhancement in tocopherols, and 1.68-fold and 1.58-fold increase in total phenolics, respectively, in comparison with the control treatment (Figure 3). Conversely, the concentrations of 100 μ M Cu and 100 μ M Cd clearly reduced nonantioxidant activity, as demonstrated by a 21% and 23% reduction in flavonols, 12% and 24% reduction in tocopherols, and 34% and 28% reduction in total phenolics, respectively, in comparison with the control treatment. We suggest that the combination of TiO_2 and EBL has a positive impact on the reduction in heavy metal toxicity by stimulating nonenzymatic antioxidant activities (flavonols, tocopherols, and total phenolics).

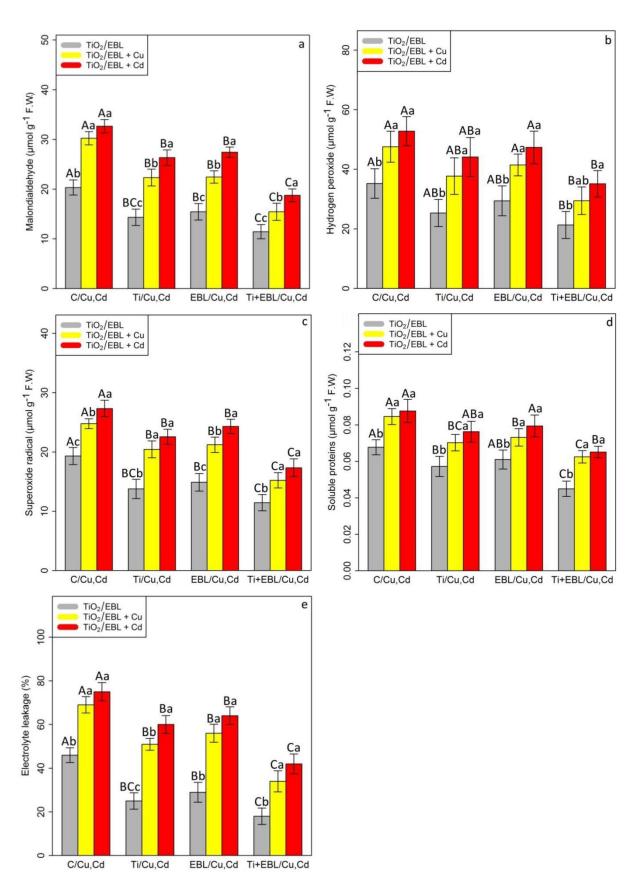


Figure 2. The impact of the co-application of 24-epibrassinolide and titanium oxide nanoparticles individually and combined on malondialdehyde content (MDA) (**a**), hydrogen peroxide (H₂O₂) (**b**), superoxide radical (O₂^{•-}) (**c**), soluble proteins (SP) (**d**), and electrolyte leakage (EL) (**e**) in bamboo species (*Pleioblastus pygmaeus*) with 100 μ M Cu and 100 μ M Cd. In this study, 1-year-old branches of

P. pygmaeus were used as plant treatments together with 100 μ M TiO₂ NPs and 10⁻⁸ M 24-epibrassinolide, individually and in combination with 100 μ M Cu and 100 μ M Cd using four replications. Planting of the treated bamboo was performed in an Air Tech inoculation hood with fluorescent white lamps and ultraviolet light (wavelengths of 10–400 nm) at 15 °C and 30 °C. The bamboo plants were constantly exposed to excess heavy metals for three weeks. Sampling for the measurement of MDA, H₂O₂, O₂•⁻, SP, and EL (**a–e**) was conducted after three weeks of plant exposure to the coapplication of 24-epibrassinolide and titanium oxide nanoparticles under 100 μ M Cu and 100 μ M Cd. The capital letters (^{A–C}) indicate significant differences between treatments of control (C), titanium (Ti), 24-epibrassinolide (EBL), and 24-epibrassinolide involving individual or combined application of titanium oxide nanoparticles (EBL–TiO₂ NPs) under 100 μ M Cu and 100 μ M Cd (the bars with similar colors), while the lowercase letters (^{a–c}) denote statistically significant differences at each concentration of the co-application of EBL and TiO₂ NPs, individually or in combination with 100 μ M Cu and 100 μ M Cd (the bars with various colors) based on Tukey's test (*p* < 0.05).

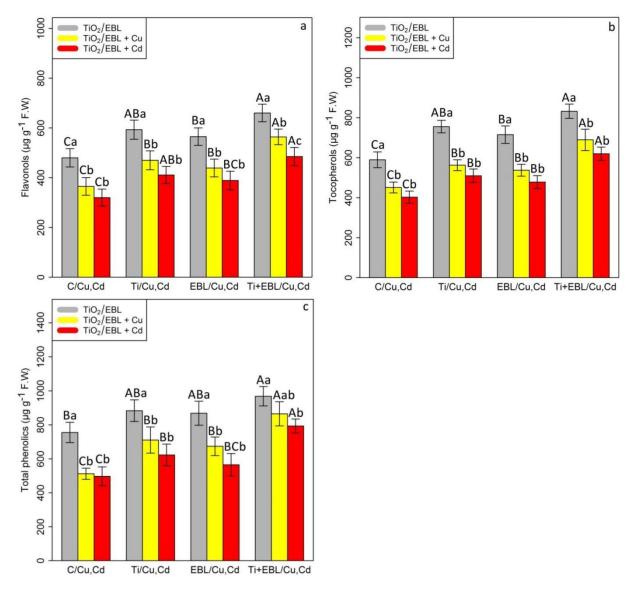


Figure 3. The impact of the co-application of 24-epibrassinolide and titanium oxide nanoparticles individually and combined on nonenzymatic antioxidant activities ((**a**) flavonols, (**b**) tocopherols, (**c**) total phenolics) in bamboo species (*Pleioblastus pygmaeus*) with 100 μ M Cu and 100 μ M Cd. In this study, 1-year-old branches of P. *pygmaeus* were used as plant treatments together with 100 μ M TiO₂ NPs and 10⁻⁸ M 24-epibrassinolide, individually and in combination with 100 μ M Cu and 100 μ M Cd,

using four replications. Planting of the treated bamboo was performed in an Air Tech inoculation hood with fluorescent white lamps and ultraviolet light (wavelengths of 10–400 nm) at 15 °C and 30 °C. The bamboo plants were constantly exposed to excess heavy metals for three weeks. Sampling for the measurement of flavonols, tocopherols, and total phenolics (**a**–**c**) was conducted after three weeks of plant exposure to the co-application of 24-epibrassinolide and titanium oxide nanoparticles under 100 μ M Cu and 100 μ M Cd. The capital letters (^{A–C}) indicate significant differences between treatments of control (C), titanium (Ti), 24-epibrassinolide (EBL), and 24-epibrassinolide involving individual or combined application of titanium oxide nanoparticles (EBL–TiO₂ NPs) under 100 μ M Cu and 100 μ M Cd (the bars with similar colors), while the lowercase letters (^{a,b}) denote statistically significant differences at each concentration of the co-application of EBL and TiO₂ NPs, individually or in combination with 100 μ M Cu and 100 μ M Cd (the bars with various colors) based on Tukey's test (*p* < 0.05).

3.4. 24-Epibrassinolide and Titanium Oxide Nanoparticles Improve Photosynthetic Pigments and Fluorescence Parameters in Bamboo Species under Cu and Cd Toxicity

Photosynthetic pigments and fluorescence parameters are important indices in the evaluation of photosynthetic efficiency in different species of plants under stress conditions. The indicators of plant photosynthesis performance, including photosynthetic pigments (Chl a, Chl b, and total Chl, as well carotenoid contents), and fluorescence indices, including the maximum photochemical efficiency of PSII (Fv/Fm), photochemical quenching coefficient (qP), effective photochemical efficiency of PSII (Fv'/Fm'), actual photochemical efficiency of PSII (φ PSII), and nonphotochemical quenching (NPQ), were measured. We found a significant difference between the co-application of 24-epibrassinolide and titanium oxide nanoparticles with Cu and Cd (p < 0.001). Based on the results, the levels of TiO₂NPs and EBL alone and in combination with heavy metals (Cu and Cd) could increase photosynthetic pigments in bamboo under Cu and Cd. However, the greatest increase in photosynthetic pigments under Cu and Cd was attributed to the co-application of TiO₂–EBL with Cu and the co-application of TiO₂–EBL with Cd, which resulted in 21% and 17% increases in Chl a, 85% and 83% increases in Chl b, 42% and 38% increases in total Chl, and 46% and 39% increases in carotenoid, respectively, in comparison with their control treatments (Table 2). Conversely, the measurement of the fluorescence parameters demonstrated a significant difference between the combination of TiO₂–EBL and Cu and Cd (p < 0.001). The data analysis revealed similar results, e.g., in the Chl and carotenoid contents and in the measurement of fluorescence parameters. Therefore, the greatest increase in fluorescence parameters was related to the combination of TiO_2 and EBL, which resulted in a 50% increase in the maximum photochemical efficiency of PSII (Fv/Fm), 41% increase in the photochemical quenching coefficient (qP), 54% increase in the effective photochemical efficiency of PSII (Fv'/Fm'), 56% increase in the actual photochemical efficiency of PSII (φ PSII), and 58% increase in nonphotochemical quenching (NPQ) in comparison with their control treatments. We suggest that the combination of TiO2 NPs and EBL has a strong ability to increase photosynthesis parameters in plants exposed to heavy metal stress (Cu and Cd) (Figure 4).

 10^{-8} M EBL + 100 μM Cu

 10^{-8} M EBL + 100 μ M Cd

 $100 \ \mu M \ TiO_2 + 10^{-8} \ M \ EBL$ $100 \ \mu M \ TiO_2 + 10^{-8} \ M \ EBL + 100 \ \mu M \ Cu$

 $100 \ \mu M \ TiO_2 + 10^{-8} \ M \ EBL + 100 \ \mu M \ Cd$

carotenoid contents) in bamboo species (<i>Pleioblastus pygmaeus</i>) with 100 μ M Cu and 100 μ M Cd.					
Treatments	Chl a (mg g $^{-1}$ F.w.)	Chl b (mg g ⁻¹ F.w.)	Chl a + b (mg g ⁻¹ F.w.)	Caratenoids (mg g $^{-1}$ F.w.)	
Control	9.64 ± 0.57 $^{ m Aa}$	7.14 ± 1.24 ^{Ba}	$16.78\pm1.76^{\text{ Ba}}$	1.35 ± 0.23 ^{Ba}	
100 μM Cu	8.51 ± 0.54 $^{ m Bab}$	$4.32\pm1.75~^{\rm Bb}$	12.83 ± 2.03 ^{Bb}	1.02 ± 0.11 ^{Bab}	
100 µM Cd	$8.35\pm0.60~^{\rm Ab}$	$3.77\pm1.16~^{\rm Bb}$	$12.12\pm1.57~^{ m Bb}$	0.92 ± 0.12 $^{ m Ab}$	
100 μM TiO ₂	10.71 ± 0.76 $^{\rm Aa}$	10.14 ± 1.57 $^{ m Aa}$	$22.35\pm3.73~^{\rm Aa}$	1.82 ± 0.11 $^{ m ABa}$	
100 μM TiO ₂ + 100 μM Cu	9.52 ± 0.67 $^{ m ABab}$	$6.54\pm0.95~^{ m ABb}$	16.06 ± 1.62 ^{Ab}	$1.32\pm0.33~^{ m ABb}$	
100 μM TiO ₂ + 100 μM Cd	$9.17\pm0.83~^{ m Ab}$	4.43 ± 1.27 ^{ABb}	$13.60\pm1.87~^{ m Bb}$	0.92 ± 0.21 ^{Ab}	
10^{-8} M EBL	$10.39\pm0.54~^{\rm Aa}$	$9.50\pm1.01~^{\rm ABa}$	$19.90\pm1.35~^{\rm Aa}$	1.50 ± 0.24 ^{Ba}	

 $5.80\pm0.97~^{ABb}$

 $4.33\pm1.31~^{ABb}$

 10.86 ± 0.82 $^{\rm Aa}$

 $8.01\pm1.68~^{\rm Ab}$

 $7.01\pm1.44~^{\rm Ab}$

 $9.09\pm0.77~^{ABab}$

 $8.85\pm0.67~^{Ab}$

 $11.05\pm1.22~^{\text{ABa}}$

 $10.23\pm1.04~^{\rm Aa}$

 9.74 ± 0.98 ABa

Table 2. The effect of the co-application of 24-epibrassinolide and titanium oxide nanoparticles individually and combined on photosynthetic pigments (Chl a, Chl b, and total Chl, as well as a) with 100 a M C

In this study, 1-year-old branches of P. pygmaeus were used as plant treatments together with 100 µM TiO₂ NPs and 10^{-8} M 24-epibrassinolide individually and in combination with 100 μ M Cu and 100 μ M Cd using four replications. Planting of the treated bamboo was performed in an Air Tech inoculation hood with fluorescent white lamps and ultraviolet light (wavelengths of 10-400 nm) at 15 °C and 30 °C. The bamboo plants were constantly exposed to excess heavy metals for three weeks. Sampling for the measurement of photosynthesis pigments was conducted after three weeks of plant exposure to the co-application of 24-epibrassinolide and titanium oxide nanoparticles under 100 μ M Cu and 100 μ M Cd. The capital letters (^{A,B}) indicate significant differences between treatments of control (C), titanium (Ti), 24-epibrassinolide (EBL), and 24-epibrassinolide with titanium oxide nanoparticles (EBL-TiO2 NPs) individually or in combination with 100 µM Cu as well as 100 µM Cd (the bars with similar colors), while the lowercase letters (a,b) denote statistically significant differences at each concentration of the co-application of EBL and TiO₂ NPs individually or in combination with 100 μ M Cu and 100 μ M Cd (the bars with various colors) based on Tukey's test (p < 0.05).

3.5. 24-Epibrasinolide and Titanium Oxide Nanoparticles Reduce Heavy Metal Accumulation in Bamboo Leaves, Stems, and Roots

The decrease in metal accumulation in different types of plants is one of the main mechanisms responsible for the reduction of metal toxicity and increase in plant resistance when exposed to oxidative stress. Our results showed that TiO_2 and EBL had a positive impact on the reduction in heavy metal concentrations in bamboo species; therefore, TiO_2 and EBL alone or in combination significantly reduced heavy metal accumulation in leaves, stems, and roots (Table 3). This phenomenon is related to the role of TiO_2 as a physical barrier that leads to a reduction in metal translocation from roots to aerial parts. The roots in plants are typically the first contact points where exposure to heavy metals occurs. Therefore, root physical traits are extremely decisive in limiting metal entry into plants. The root-based cellular layers comprised of epiblema, endodermis, and exodermis form apoplastic barriers in the roots, which can restrict heavy metal uptake by plants. We hypothesized that TiO_2 NPs, through strengthening the apoplastic barriers in the roots and enhancing their impermeability, can significantly diminish the uptake of heavy metals. On the other hand, TiO_2 NPs with high adsorption capacity act as an efficient binder of metal ions. Hence, TiO₂ NPs can restrain the movement of heavy metals within the extracellular or intercellular parts of roots, thereby restricting heavy metal translocation from the root to shoot. TiO₂ NPs may also have the ability to influence the expansion of the epidermal layer in plants, preventing heavy metal accumulation in nonphotosynthetic tissues by providing additional physical resistance. Conversely, we indicated that EBL, a hormone that is involved in plant growth regulation alone and in combination with TiO_2 , plays a positive role in the stimulation of antioxidant activity, which can scavenge ROS components in plant organs and inhibit plant oxidative stress caused by heavy metal toxicity. As shown in Table 3, the combination of TiO_2 –EBL with heavy metals showed the greatest reduction in heavy metal accumulation in the leaves, stems, and roots of bamboo species (Table 3). We suggest that TiO₂ and EBL can reduce heavy metal contents in plant leaves, stems, and

 $1.09\pm0.09~^{ABa}$

 1.09 ± 0.25 Aa

 $2.27\pm0.27~^{Aa}$

 $1.48\pm0.08~^{\rm Ab}$

 1.30 ± 0.12 Ab

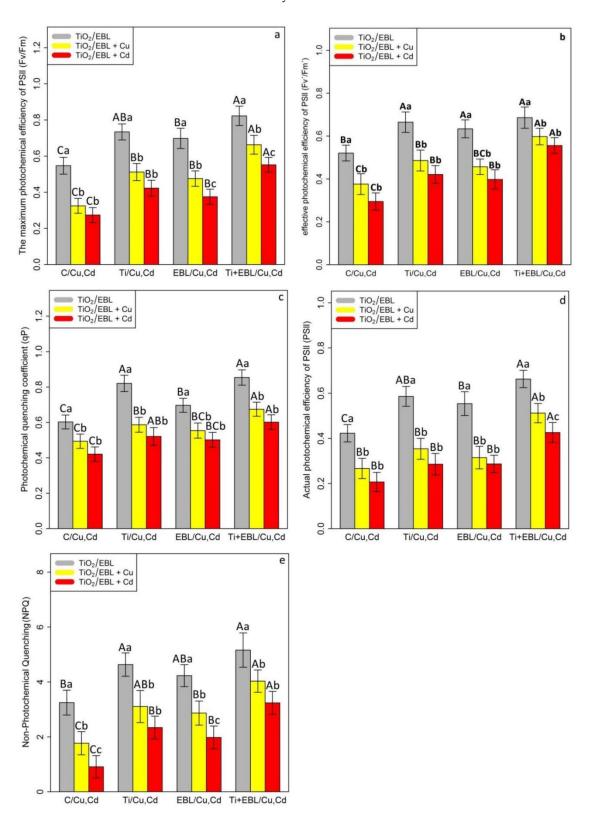
 $14.89\pm1.66\ ^{Ab}$

 $13.18\pm1.97~^{Ab}$

 $21.92\pm1.97~^{Aa}$

 $18.24\pm1.72~^{\rm Aab}$

 16.75 ± 2.08 Ab



roots, thus demonstrating that the combination of TiO_2 and EBL has the greatest impact on the decrease in metal toxicity.

Figure 4. The effect of the co-application of 24-epibrassinolide and titanium oxide nanoparticles individually and combined on fluorescence parameters, including the maximum photochemical efficiency of PSII (Fv/Fm) (**a**), effective photochemical efficiency of PSII (Fv'/Fm') (**b**), photochemical quenching coefficient (qP) (**c**), actual photochemical efficiency of PSII (ϕ PSII (ϕ), and nonphotochemical quenching

(NPQ) (e) in bamboo species (*Pleioblastus pygmaeus*) with 100 μ M Cu and 100 μ M Cd. In this study, 1-year-old branches of *P. pygmaeus* were used as plant treatments together with 100 μ M Ci₂ NPs and 10⁻⁸ M 24-epibrassinolide individually and in combination with 100 μ M Cu and 100 μ M Cd using four replications. Planting of the treated bamboo was performed in an Air Tech inoculation hood with fluorescent white lamps and ultraviolet light (wavelengths of 10–400 nm) at 15 °C and 30 °C. The bamboo plants were constantly exposed to excess heavy metals for three weeks. Sampling for the measurement of fluorescence parameters (**a**–**e**) was conducted after three weeks of plant exposure to the co-application of 24-epibrassinolide and titanium oxide nanoparticles under 100 μ M Cu and 100 μ M Cd. The capital letters (^{A–C}) indicate significant differences between treatments of control (C), titanium (Ti), 24-epibrassinolide (EBL), and 24-epibrassinolide involving individual or combined application of titanium oxide nanoparticles (EBL–TiO₂ NPs) under 100 μ M Cu and 100 μ M Cd (the bars with similar colors), while the lowercase letters (^{a–c}) MPs individually or in combination with 100 μ M Cu and 100 μ M Cd (the bars with various colors) based on Tukey's test (*p* < 0.05).

Table 3. The accumulation concentrations of titanium oxide nanoparticles and corresponding heavy metals (Cu and Cd) in bamboo shoots, stems, and roots.

Treatments	Heavy Metal in Leaves (µmol L ⁻¹)	TiO ₂ NP in Leaves (μmol L ⁻¹)	Heavy Metal in Stem (µmol L ⁻¹)	TiO ₂ NP in Stem (μmol L ⁻¹)	Heavy Metal in Root (µmol L ⁻¹)	TiO ₂ NP in Root (μmol L ⁻¹)
Control	0	0	0	0	0	0
100 µM Cu	19.22 ± 1.10 ^{Ab}	0	25.3 ± 1.03 $^{ m Ab}$	0	31.2 ± 1.00 ^{Ab}	0
100 µM Cd	$24.20\pm1.12~^{\rm Aa}$	0	$29.8\pm1.02~^{\rm Aa}$	0	36.3 ± 1.02 $^{\mathrm{Aa}}$	0
100 µM TiO ₂	0	$18.5\pm0.90~^{\rm Aa}$	0	$24.5\pm0.95~^{\rm Aa}$	0	$34.6\pm1.04~^{\rm Aa}$
100 μM TiO ₂ + 100 μM Cu	$13.42\pm1.10~^{\rm Bb}$	14.3 ± 0.79 ^{Ab}	$16.6\pm0.90~^{\rm Cb}$	18.5 ± 0.86 $^{ m Ab}$	$19.1\pm1.11~^{\mathrm{Cb}}$	$28.5\pm0.85~^{\rm Ab}$
100 μM TiO ₂ + 100 μM Cd	16.80 ± 0.97 ^{Ba}	$10.8\pm0.87~^{ m Ac}$	$21.2\pm0.86~^{\rm Ca}$	12.3 ± 0.86 $^{ m Ac}$	25.4 ± 0.95 ^{Ca}	$20.5\pm0.90~^{\rm Ac}$
10^{-8} M EBL	0	0	0	0	0	0
10 ⁻⁸ M EBL + 100 μM Cu	14.60 ± 1.07 ^{Bb}	0	18.8 ± 0.98 ^{Bb}	0	22.3 ± 0.94 ^{Bb}	0
10 ⁻⁸ M EBL +100 µM Cd	17.41 ± 0.97 ^{Ba}	0	23.4 ± 0.94 $^{\mathrm{Ba}}$	0	28.6 ± 0.90 ^{Ba}	0
$100 \ \mu M \ TiO_2 + 10^{-8} \ M \ EBL$	0	16.4 ± 0.77 $^{\mathrm{Ba}}$	0	$21.3\pm0.95~^{\text{Ba}}$	0	$31.3\pm0.94~^{Ba}$
100 μM TiO ₂ + 10 ⁻⁸ M EBL +100 μM Cu	$8.40\pm1.07~^{\rm Cb}$	$11.2\pm0.87~^{\text{Bb}}$	$12.4\pm1.02~^{\text{Db}}$	$15.4\pm0.95~^{\text{Bb}}$	$15.4\pm0.94~^{\text{Db}}$	$24.5\pm1.02~^{Bb}$
100 μM TiO ₂ + 10 ⁻⁸ M EBL +100 μM Cd	$10.36\pm0.99~^{\rm Ca}$	$8.3\pm0.90~^{Bc}$	$15.4\pm0.97^{\text{ Da}}$	$10.2\pm0.94~^{\text{Bc}}$	$17.7\pm0.94~^{\rm Da}$	$16.4\pm0.86~^{\rm Bc}$

In this study, 1-year-old branches of *P. pygmaeus* were used as plant treatments together with 100 μ M TiO₂ NPs and 10⁻⁸ M 24-epibrassinolide individually and combined with 100 μ M Cu and 100 μ M Cd using four replications. The capital letters (^{A–D}) indicate significant differences between treatments of control (C), titanium (Ti), 24-epibrassinolide (EBL), and 24-epibrassinolide with titanium oxide nanoparticles (EBL–TiO₂ NPs) individually or in combination with 100 μ M Cu as well as 100 μ M Cd (the bars with similar colors), while the lowercase letters (^{a–c}) denote statistically significant differences at each concentration of the co-application of EBL and TiO₂ NPs individually or in combination with 100 μ M Cu and 100 μ M Cd (the bars with various colors) based on Tukey's test (*p* < 0.05). 2–6 24-Epibrasinolide and titanium oxide nanoparticles increase plant biomass indices (root and shoot dry weight) and plant growth (length of shoot) in bamboo species under Cu and Cd toxicity.

To evaluate the plant growth rate under Cu and Cd toxicity, the plant biomass, including root and shoot dry weight, as well as the length of shoots, were measured. A significant difference was found for the co-application of TiO₂–EBL and heavy metals (p < 0.001) (Figure 5). Therefore, TiO₂ and EBL individually and in combination significantly increased plant growth and biomass under stressful conditions. Based on this result, the greatest increase in plant biomass and growth under heavy metal exposure was related to the combination of TiO₂ and EBL, which resulted in a 21% increase in the dry weight of shoots, a 23% increase in the dry weight of roots, and a 19% increase in the length of shoots in comparison with their control treatments (Table 4). Conversely, the measurements showed that the lowest plant growth was recorded under 100 μ M Cu and 100 μ M Cd, which resulted in 0.5 g and 0.46 g dry weights of shoots, 0.59 g and 0.53 g dry weights of roots, and 10.04 cm and 9.21 cm shoot lengths, respectively (Table 4).

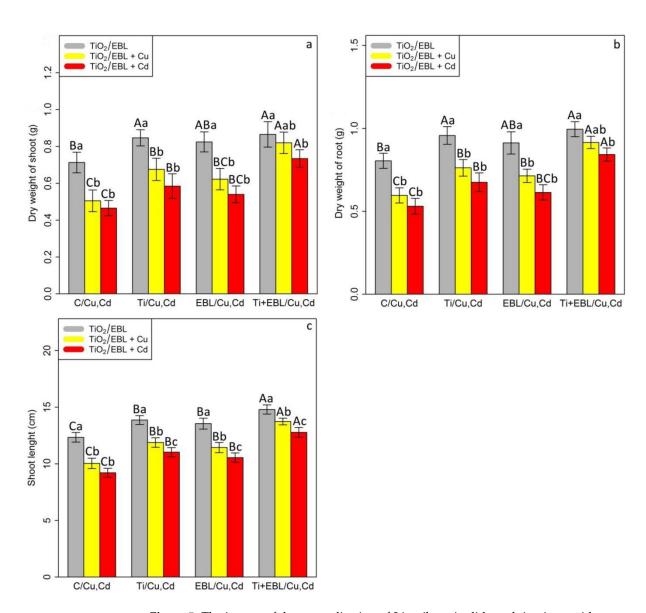


Figure 5. The impact of the co-application of 24-epibrassinolide and titanium oxide nanoparticles individually and combined on the dry weight of shoots (a), dry weight of roots (b), and shoot length (c) of bamboo species (Pleioblastus pygmaeus) with 100 µM Cu and 100 µM Cd. In this study, 1-year-old branches of P. pygmaeus were used as plant treatments together with 100 µM TiO₂ NPs and 10⁻⁸ M 24epibrassinolide individually and combined with 100 µM Cu and 100 µM Cd through four replications. Planting of the treated bamboo was performed in an Air Tech inoculation hood with fluorescent white lamps and ultraviolet light (wavelengths of 10-400 nm) at 15 °C and 30 °C. The bamboo plants were constantly exposed to excess heavy metals for three weeks. Sampling for the measurement of biomass indexes and plant growth (a-c) was conducted after three weeks of plant exposure to the co-application of 24-epibrassinolide and titanium oxide nanoparticles under 100 µM Cu and 100 μ M Cd. The capital letters (^{A-C}) indicate significant differences between treatments of control (C), titanium (Ti), 24-epibrassinolide (EBL), and 24-epibrassinolide involving individual or combined application of titanium oxide nanoparticles (EBL-TiO₂ NPs) under 100 μ M Cu and 100 μ M Cd (the bars with similar colors), while the lowercase letters (^{a-c}) denote statistically significant differences at each concentration of the co-application of EBL and TiO₂ NPs individually or in combination with 100 μ M Cu and 100 μ M Cd (the bars with various colors) based on Tukey's test (p < 0.05).

Treatments	Dry Weight of Shoot (%)	Dry Weight of Root (%)	Shoot Length (%)
100 μM Cu	29%↓	26%↓	18%↓
100 µM Cd	35%↓	34%↓	25%↓
100 μM TiO ₂	$18\%\uparrow$	19%↑	12%↑
100 μM TiO ₂ + 100 μM Cu	5%↓	5%↓	3%↓
100 μM TiO ₂ + 100 μM Cd	17%↓	16%↓	10%↓
10^{-8} M EBL	15%↑	13%↑	$10\%\uparrow$
10^{-8} M EBL + 100 μ M Cu	12%↓	11%↓	7%↓
10^{-8} M EBL + 100 μ M Cd	24%↓	23%↓	14%↓
$100 \ \mu M \ TiO_2 + 10^{-8} \ M \ EBL$	21%↑	23%↑	19%↑
$100 \ \mu M \ TiO_2 + 10^{-8} \ M \ EBL + 100 \ \mu M \ Cu$	15%↑	13%↑	11%↑
$100 \mu\text{M}\text{Ti}\text{O}_2 + 10^{-8}\text{M}\text{EBL} + 100\mu\text{M}\text{Cd}$	3%↑	$4\%\uparrow$	3%↑

Table 4. The changes in bamboo biomass in root and shoot dry weight as well as shoot length with 24-epibrassinolide and titanium oxide nanoparticles individually or combined with 100 μ M Cu and 100 μ M Cd in comparison with the control treatment.

3–7 24-Epibrasinolide and titanium oxide nanoparticles reduce the translocation factor (TF) and bioaccumulation factor (BAF) and improve the tolerance index (TI) in roots and shoots of bamboo species. \uparrow indicates increases and \downarrow indicates decreases.

The translocation factor (TF) is one of the main mechanisms used to evaluate the remediation efficiency of heavy metals in plant organs and the reduction of toxicity in plants under heavy metal stress. Therefore, it is calculated according to differences in the accumulation of Cu and Cd in shoots and roots, and it serves as an important factor in increasing plant tolerance to toxicity. In the present study, the addition of 24-epibrassinolide and titanium oxide nanoparticles significantly reduced Cu and Cd translocation from roots to shoots, which led to a reduction in toxicity by limiting metal accumulation in the plant aerial organs. Therefore, according to the results, the co-application of 24-epibrassinolide and titanium oxide nanoparticles in combination with heavy metals (Cu and Cd) resulted in a low level of translocation factor, which could reduce metal toxicity in the aerial parts of bamboo plants (Table 5). Additionally, the result showed that the co-application of EBL and TiO₂ NPs significantly reduced Cu and Cd concentration in the leaves (p < 0.001), implicating the positive role of EBL-TiO₂NPs in the reduction of heavy metal toxicity in the aerial parts of the bamboo plant (Table 5). Conversely, the calculation of the tolerance indices in shoots and roots revealed a significant difference between the co-application of TiO₂ NPs and EBL alone under Cu and Cd (p < 0.001). Therefore, the levels of TiO₂ NPs and EBL indicated an increase in shoot and root tolerance under heavy metal stress, which was obtained by the amelioration mechanism of the co-application of TiO_2 and EBL against heavy metal toxicity, such as the stimulation of antioxidant activity and the increase in plant biomass. We suggest that TiO₂ NP and EBL concentrations alone increase plant tolerance under metal stress; however, the most positive effect was more pronounced with the co-application of TiO_2 NPs and EBL under Cu and Cd toxicity (Table 5).

Table 5. Changes in the translocation factor and tolerance index of shoots and roots in response to 24-epibrassinolide and titanium oxide nanoparticles individually or in combination with 100 μ M Cu and 100 μ M Cd compared with the control treatment. Each data point is the mean \pm SE of four replicates. The capital letters (^{A–C}) indicate significant differences between treatments of control (C), titanium (Ti), 24-epibrassinolide (EBL), and 24-epibrassinolide involving individual or combined application of titanium oxide nanoparticles (EBL–TiO₂ NPs) under 100 μ M Cu and 100 μ M Cd (the bars with similar colors), while the lowercase letters (^{a–c}) denote statistically significant differences at each concentration of the co-application of EBL and TiO₂ NPs individually or in combination with 100 μ M Cu and 100 μ M Cd (the bars with various colors) based on Tukey's test (*p* < 0.05).

Treatment	Translocation Factor (Leaves)	Tolerance Index (Shoot)	Tolerance Index (Root)	Bioaccumulation Factor (Leaves)
Control	$0.00\pm0.00~^{\rm Bb}$	$1.00\pm0.00~^{\rm Ba}$	$1.00\pm0.00~^{\rm Ba}$	$0.00\pm0.00~^{\rm Aa}$
100 µM Cu	0.65 ± 0.02 $^{\mathrm{Aa}}$	0.70 ± 0.06 ^{Cb}	0.74 ± 0.09 ^{Bb}	0.19 ± 0.01 ^{Ab}
100 µM Cd	$0.66\pm0.01~^{\rm Aa}$	0.65 ± 0.05 ^{Cb}	0.66 ± 0.09 ^{Bb}	$0.24\pm0.01~^{ m Ac}$
100 μM TiO ₂	0.52 ± 0.04 $^{ m Ab}$	$1.19\pm0.13~^{ m ABa}$	1.23 ± 0.12 $^{ m Aa}$	0.00 ± 0.00 Aa
100 μM TiO ₂ + 100 μM Cu	$0.58\pm0.01~^{\mathrm{BCa}}$	$0.94\pm0.07~^{ m ABb}$	$0.95\pm0.11~^{ m ABb}$	$0.13\pm0.01~^{\text{Bb}}$
100 μM TiO ₂ + 100 μM Cd	$0.60\pm0.01~^{\rm ABa}$	0.80 ± 0.05 ^{Bb}	0.84 ± 0.10 ^{Bb}	$0.16\pm0.01~^{\rm Bc}$
10^{-8} M EBL	$0.00\pm0.00~^{ m Bb}$	$1.15\pm0.09~^{ m ABa}$	1.13 ± 0.13 $^{ m ABa}$	0.00 ± 0.00 Aa
10 ⁻⁸ M EBL + 100 μM Cu	0.60 ± 0.02 ^{Ba}	$0.87\pm0.08~^{ m BCb}$	$0.89\pm0.09~^{ m ABb}$	$0.14\pm0.01~^{\rm Bb}$
10 ⁻⁸ M EBL + 100 μM Cd	$0.61\pm0.05~^{\rm Aa}$	$0.75\pm0.01~^{\rm Bb}$	0.76 ± 0.09 ^{Bb}	$0.17\pm0.00~^{\rm Bc}$
100 μM TiO ₂ + 10^{-8} M EBL	0.49 ± 0.01 ^{Ab}	$1.21\pm0.08~^{\rm Aa}$	1.24 ± 0.12 $^{ m Aa}$	0.00 ± 0.00 Aa
100 μM TiO ₂ + 10^{-8} M EBL + 100 μM Cu	$0.53\pm0.02~^{\rm Cab}$	1.1 ± 0.09 ^{Aab}	$1.09\pm0.10~^{\rm Aa}$	$0.08\pm0.01~^{\rm Cb}$
$100 \ \mu M \ TiO_2 + 10^{-8} \ M \ EBL + 100 \ \mu M \ Cd$	$0.54\pm0.02~^{\rm Ba}$	$1.02\pm0.04~^{\rm Ab}$	$1.04\pm0.08~^{\rm Aa}$	$0.10\pm0.01~^{\rm Bc}$

4. Discussion

Titanium, as a form of TiO_2 , has the ability to alter the bioavailability and behavior of metals in the environment [45]. The impact of TiO₂ NPs on increasing antioxidant activity and plant growth has been reported in several studies [13,14,46]. This increase can be attributed to the inductive role of TiO₂ NPs in enhancing signaling associated with the activation of antioxidant enzyme activity [13]. This finding is consistent with the reported results in the present study. Therefore, our results demonstrated that the individual levels of TiO₂ NPs could increase antioxidant and nonantioxidant activity in bamboo plants under certain Cu and Cd levels. Conversely, the level of EBL regulates plant stress by stimulating antioxidant activity [47]. The increasing capacity of antioxidant activity based on the levels of EBL in plants under stress has been reported in many studies [48–50]. EBL seems to play a main role in the activation of genes responsible for antioxidants by stimulating the expression of genes responsible for SOD, CAT, and APX in plants exposed to heavy metal stress [51]. Hence, it is interesting to note that EBL has the ability to ameliorate oxidative stress caused by metals, which has previously been reported for many plant species, such as Brassica juncea [20], Cicer arietinum [52], and Raphanus sativa [53]. The main reason can be attributed to the role of BR signaling kinase (BSK 1) in the stimulation of salicylic acid levels against oxidative damage [54]. In our studies, the application of TiO_2 and EBL individually and in combination enhanced antioxidant enzyme activities, including SOD, POD, CAT, GR, APX, and PAL. However, the combination of TiO₂ NP and EBL was more effective in increasing antioxidant levels than TiO₂ NPs and EBL alone. Phenolic compounds, as nonenzymatic antioxidant activities, alleviate the negative effect of reactive oxygen radicals and have a strong ability to chelate metals [55,56]. There seems to be a relationship between enhancing phenylalanine ammonia-lyase (PAL) and the total phenolic compound, and it has been reported that PAL is a key enzyme responsible for the activation of the synthesis of phenolic compounds under stress [57,58]. This phenomenon has been reported in some studies on the reduction of Cu and Cd toxicity [55,59]. Our results demonstrated that the application of TiO₂ NPs and EBL individually and in combination increased nonantioxidant activity (total phenolics, flavonols, and tocopherols) under Cu and Cd toxicity. This phenomenon could be related to PAL gene transcript levels as well as increasing PAL activity in response to EBL levels under heavy metal stress, which ultimately reduces ROS compounds by synthesizing phenolic compounds.

When antioxidant activity increases, the plant experiences cellular injuries (H_2O_2 , MDA, and EL) [60]. It has been reported that TiO_2 NPs induce certain stress-combative mechanisms, such as an improvement of the defense mechanism against ROS accumulation in plant intercellular space, which has been shown to attenuate H_2O_2 induction [60]. An increase in MDA content has been described as the initial stage of plant injury, which shows the rate of membrane lipid peroxidation [61]. This reveals the extent to which plants face this serious problem. Based on the present findings, we suggest that TiO₂ NPs protect the plant cell membrane from ROS, a phenomenon that has been related to the role of TiO₂ NPs in boosting antioxidant activity. Conversely, EBL has the ability to positively alter the membrane structure and membrane stability in plants exposed to stresses, such as heavy metals, which leads to a reduction in membrane lipid peroxidation [62]. In one study, the level of EBL was observed to diminish the concentrations of H_2O_2 and MDA (20-60% reductions) in plants under Pb stress [63]. In another study, EBL diminished the oxidative toxicity in cowpea under Cd by reducing lipid peroxidation, MDA content, and electrolytes [64,65]. In the present study, the level of EBL improved the ROS content and reduced the plant cell membrane under heavy metal toxicity. It is interesting to note that in this study, the application of TiO_2 NPs and EBL individually and in combination decreased ROS and lipid peroxidation, including H₂O₂, O₂^{•-}, MDA, SP, and EL, in bamboo plants exposed to metal stress. One of the mechanisms underlying the amelioration of lipid peroxidation and ROS by EBL can be attributed to the increase in endogenous plant hormones that regulate plant growth, such as salicylic acid and ethylene, and the cross-talk between them. These mechanisms can improve plant tolerance under metal toxicity [66]. In this study, the co-application of TiO_2 NPs and EBL was more efficient in reducing ROS compounds and ameliorating lipid peroxidation than TiO₂ NPs and EBL alone.

Studies have indicated that TiO₂ NPs enhance photosystem II in spinach by promoting oxygen evolution and energy transfer [67]. Additionally, TiO₂ reduces Chl degradation and stimulates Chl biosynthesis, which can promote photosynthesis by stabilizing chlorophylls and carotenoids [60]. Chlorophylls are the most abundant component of the chloroplast and play an efficient role in the rate of photosynthesis [59]. The role of EBL in enhancing the cell number and photosynthetic pigment content (Chl a, Chl b, and carotenoids) has been demonstrated in some studies [68,69]. It seems that the increase in photosynthesis and Chl pigments by EBL is related to the stimulation activity of ribulose 1,5-bisphosphate carboxylase oxygenase as well as the increase in the Calvin cycle enzymes [20]. Therefore, the level of EBL with increasing carotenoids ameliorates photodamage during photosynthesis [70]. According to the above mechanisms, the co-application of EBL and Ti can improve photosynthesis and Chl pigment levels via Chl biosynthesis and reduce photodamage via the activation of the Calvin cycle enzymes in plants under heavy metal stress. Additionally, the results revealed a positive impact of the co-application of EBL and TiO₂ NPs on fluorescence parameters, which showed an increase in the efficiency of fluorescence indices, including the maximum photochemical efficiency of PSII (Fv/Fm), photochemical quenching coefficient (qP), effective photochemical efficiency of PSII (Fv'/Fm'), actual photochemical efficiency of PSII (φ PSII), and nonphotochemical quenching (NPQ). Therefore, we suggest that the co-application of EBL and TiO_2 NPs can increase photosynthetic properties in plants exposed to heavy metals (Cu and Cd), and this phenomenon could be related to the increase in antioxidant activity and the reduction in heavy metal accumulation under metal toxicity stress.

Titanium is known to be the most abundant transition element after iron, with a level of 1–578 mg kg⁻¹ in different species of non-hyperaccumulator plants [71]. However, low mobility in the soil may impact its absorption by plants [72]. Our results showed that the root accumulation of TiO₂ NPs was higher than that of stems and shoots, indicating that bamboo roots prefer to be storage organs of titanium, which has been reported in another study [73]. Therefore, the accumulation of titanium NPs led to the adsorption of heavy

metals in roots, which, as a physical barrier, reduces metal translocation from roots to shoots. Conversely, EBL has the potential to reduce metal accumulation in plants by enhancing phytochelatin synthesis (PC) [48-50]. In a study on sugar beet, the levels of EBL reduced the absorption of heavy metals by plants by 50% [74–76]. It has also been reported [69] to reduce Cd accumulation in the roots, stems, and shoots of pea seedlings. Conversely, EBL levels reduce the accumulation of Cd by preserving ion homeostasis through the acceleration of calcium absorption [77,78]. Therefore, EBL leads to increased absorption of K⁺, mg⁺, and K⁺, which can be transported to the aerial parts of plants, such as leaves, and finally limit Cd and metal translocation from roots to shoots [79]. In the present study, the application of EBL and TiO₂ could individually and in combination diminish the accumulation of heavy metals. These results are related to the role of EBL in preserving ion homeostasis, which can limit heavy metal uptake by coperception, which is associated with the role of TiO_2 in the adsorption and absorption of heavy metals on the root surface. Similar to other transition metals, titanium, present in small fractions, absorbs to and accumulates in roots, as it is translocated through the xylem stream from roots to shoots [80]. The translocation of TiO₂ NPs to aerial plant parts has been demonstrated [81,82]. As shown in Table 5, the co-application of TiO₂ NPs and EBL significantly reduced metal translocation from roots to shoots, which is an important mechanism in increasing the tolerance of bamboo plants. However, the results showed that BAF in the roots was higher than in the stem and the leaves, which indicated that TiO₂ NPs–EBL could effectively reduce BAF in the aerial parts of the bamboo plant. This could be explained by the mechanisms involved in the absorbance of Cu and Cd in the root surface by the co-application of TiO2 NPs and EBL. Therefore, we suggest that TiO_2 NPs–EBL has an important role in the reduction of adsorption and uptake as well as the translocation of Cu and Cd to the aerial parts (leaves and stem). Thus, the application of TiO_2 NPs–EBL can retain the heavy metals on the bamboo root surface. On the other hand, the results showed that the TF in the leaves was less than that in the stem, which was an indication that the heavy metals had been accumulated in the leaves less than in the stem (Figure 6).

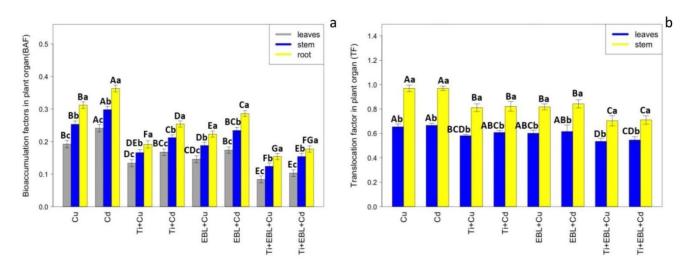


Figure 6. The impact of the co-application of 24-epibrassinolide and titanium oxide nanoparticles on BAF and TF: (**a**) the comparison between bioaccumulation of heavy metals in the root, stem, and leaves; (**b**) the comparison between translocation factor in the stem and the leaves. Bioaccumulation factor (BAF) is obtained by the difference between the concentrations of heavy metals in the leaf, stem, or root and concentrations of heavy metals in the medium, while the translocation factor (TF) is obtained by the difference between the concentration factor (Cd) in the leaves or stem of plants and the concentration of the heavy metals (Cu, Cd) in the roots of plants. The capital letters (^{A–G}) indicate significant differences between treatments of control (C), titanium (Ti), 24-epibrassinolide (EBL), and 24-epibrassinolide involving individual or combined application of titanium oxide nanoparticles (EBL

 $-\text{TiO}_2$ NPs) under 100 μ M Cu and 100 μ M Cd (the bars with similar colors), while the lowercase letters (^{a-c}) denote statistically significant differences between leaves, stem, and root at each treatment (the bars with various colors) based on Tukey's test (p < 0.05).

Titanium is a useful element that can aid plant growth by increasing plant photosynthesis and enzymatic activity as well as increasing plant uptake of other nutrients [72]. One study reported that TiO₂ can enhance the absorption rate of micro-and macronutrients, which could be the main factor in plant growth and biomass [83]. In our study, the role of TiO_2 in increasing the plant biomass was related to an increase in nutrient absorbance by the plants and reduced toxicity in response to an increase in antioxidant activity. Many studies have reported that EBL increases plant growth under heavy metal stress [21,84,85]. The reduction in plant growth by heavy metal toxicity is related to the number of intercellular metal ions bound to the surface of the cell [86,87]. Our results showed that EBL could increase the plant biomass and plant growth under heavy metal toxicity, which could be related to the role of EBL in the reduction in intercellular metal ions, a phenomenon that has been confirmed in a study on A. obliquus [63]. However, the role of EBL in plant growth regulation during the stimulation of plant defense mechanisms can also be considered. Therefore, we hypothesized that EBL increased the plant growth under heavy metal stress by promoting antioxidant capacity. The increase in Chl as well as carotenoid contents in response to EBL has been confirmed in many studies [88–91]. Conversely, EBL has the ability to control cell division and elongation by regulating xyloglucan endotransglucosylase [92,93], thus demonstrating the positive role of EBL on plant growth and development, especially under stressful conditions. In the present research, the role of the co-application of TiO₂ NPs and EBL in improving plant biomass and plant growth seemed to be related to the ameliorative mechanisms activated by the levels of both TiO_2 NPs and EBL. The application of small-sized nanoparticles in the range of 1–100 nm is a new strategy to maintain plant growth and development under heavy metals and other abiotic stresses. However, the build-up of TiO_2 NPs within the plant organs can have dual effects of growth promotion and suppression. Titanium dioxide nanoparticles (TiO₂ NPs) lead to several beneficial outcomes on the physiological, morphological, and biochemical traits of some plant species, which has been indicated in some studies [94,95] as well as our present study. Conversely, some researchers have reported the detrimental impacts of high levels of TiO_2 NPs on plants [96,97]. These implications might arise due to various environmental conditions, different plant species, and the applied levels [97,98]. Therefore, the safety/danger of TiO₂ NPs for plants depends on a myriad of factors including size, concentration, method of treatment application, plant type and growth pattern, uptake amount by plants, cellular chemical properties, translocation rate, and reactivity of TiO₂ NPs in various tissues, which determine NP interaction with a wide array of metabolic activities of the plants that can thus lead to their advantageous or toxic effects [99,100]. Additionally, the TiO_2 NP surface area, their predisposition for accumulation in tissues, and their inherent reactivity are the possible reasons for their toxicological repercussions [101]. Therefore, there is a great need to take the aforementioned variables into careful consideration while applying TiO_2 NPs in the agriculture and food industry, which can minimize health risk for humans. This finding revealed the effective role of the co-application of TiO_2 NPs and EBL in comparison with TiO₂ NPs and EBL individually.

5. Conclusions

Heavy metals are deemed a considerable environmental safety hazard with inhibitory impacts on plant growth due to the induction of excessive levels of ROS compounds, which causes oxidative stress in cells and tissues. The use of nanoparticles and phytohormones as two possible agents that mitigate the deleterious effects of heavy metals has been on the rise in the recent years. Thus, conducting extensive research using various plant species with distinct growth and morphological characteristics is needed. Based on our experimental results, the individual application of EBL as a phytohormone and TiO_2 NPs contributed

to the amelioration of toxicity in bamboo plants under excess Cu and Cd. However, the co-application of EBL and TiO_2 demonstrated a greater effective influence on increasing plant tolerance under metal toxicity. Therefore, our results indicated that while Cu and Cd stress led to increased ROS production, causing injury to the plant membrane, by boosting oxidative activity, the co-application of EBL and TiO_2 significantly reduced the ROS content and oxidative stress in the plants, resulting in an increase in the photosynthetic properties and an enhancement in the plant growth and development. Conversely, the co-application of EBL and TiO_2 increased the plant tolerance under metal toxicity by reducing the heavy metal accumulation within the plant and restricting the metal translocation from the roots to the shoots. Overall, our study revealed the cellular-and tissue-level mechanisms involved in increasing bamboo plant tolerance to Cu and Cd toxicity through the integrated use of EBL and TiO_2 . This result requires further investigation with different plant species.

Author Contributions: Conceptualization, A.E., Y.D. and J.B.; statistical analysis, A.E. and Y.L.; Investigation, A.E. and G.L.; Supervision, A.E., Y.D., and G.L.; Project Administration, A.E.; Funding Acquisition, A.E. and G.L.; writing—original draft and revised preparation, A.E., Y.D., J.B., F.M., M.R., and G.L.; writing—review and editing, J.B., F.M., and M.H.; visualization, M.H. All authors have read and agreed to the published version of the manuscript.

Funding: This work received financial support from Nanjing Forestry University (Start-Up Research Fund) and the Bamboo Research Institute. Special Funding for this work was provided by Jiangsu Agricultural Science and Technology Innovation Fund, No. CX (18) 2031.

Institutional Review Board Statement: The study was conducted according to the guidelines of the Declaration of Helsinki and approved by the Institutional Review Board of Nanjing Forestry University.

Informed Consent Statement: Not applicable.

Data Availability Statement: The data presented in this study are available in article.

Acknowledgments: We would like to extend our sincere gratitude and appreciation to Peijian Shi, Co-Innovation Center for Sustainable Forestry in Southern China, Nanjing Forestry University, Nanjing, Jiangsu, China, for helping in the statistical analysis of the manuscript.

Conflicts of Interest: The authors declare that there is no conflict of interests regarding the publication of this paper.

References

- Bhat, J.A.; Shivaraj, S.M.; Singh, P.; Navadagi, D.B.; Tripathi, D.K.; Dash, P.K.; Solanke, A.U.; Sonah, H.; Deshmukh, R. Role of silicon in mitigation of heavy metal stresses in crop plants. *Plants* 2019, *8*, 71. [CrossRef]
- Kim, J.J.; Kim, Y.S.; Kumar, V. Heavy metal toxicity: An update of chelating therapeutic strategies. J. Trace Elem. Med. Biol. 2019, 54, 226–231. [CrossRef] [PubMed]
- 3. Qi, Z.Y.; Ahammed, G.J.; Jiang, C.Y.; Li, C.X.; Zhou, J. E3 ubiquitin ligase gene SlRING1 is essential for plant tolerance to cadmium stress in *Solanum lycopersicum*. J. Biotechnol. 2020, 324, 239–247. [CrossRef] [PubMed]
- 4. Zhang, X.; Zhong, T.; Liu, L.; Ouyang, X. Impact of soil heavy metal pollution on food Safety in China. *PLoS ONE* 2015, 10, e0135182. [CrossRef] [PubMed]
- Adrees, M.; Ali, S.; Rizwan, M.; Zia-ur-Rehman, M.; Ibrahim, M.; Abbas, F.; Farid, M.; Qayyum, M.F.; Irshad, M.K. Mechanisms of silicon-mediated alleviation of heavy metal toxicity in plants: A review. *Ecotoxicol. Environ. Saf.* 2015, 119, 186–197. [CrossRef]
- 6. Yruela, I. Copper in plants. Braz. J. Plant Physiol. 2015, 17, 145–156. [CrossRef]
- 7. Bouazizi, H.; Jouili, H.; Geitmann, A.; El Ferjani, E. Copper toxicity in expanding leaves of *Phaseolus vulgaris* L.: Antioxidant enzyme response and nutrient element uptake. *Ecotoxicol. Environ. Saf.* **2010**, *73*, 1304–1308. [CrossRef]
- 8. Jouilia, H.; Bouazizia, H.; Rossignol, M.; Borderies, G.; Jamet, E.; El Ferjani, E. Partial purification and characterization of a copper-induced anionic peroxidase of sunflower roots. *Plant Physiol. Biochem.* **2008**, *46*, 760–767. [CrossRef]
- 9. Singh, S.; Susan, E.; D'Souza, S.F. Cadmium accumulation and its influence on lipid peroxidation and antioxidative system in an aquatic plant, *Bacopa monnieri* L. *Chemosphere* **2006**, *62*, 233–246. [CrossRef]
- Li, M.; Ahammed, G.J.; Li, C.; Bao, X.; Yu, J.; Huang, C.; Yin, H.; Zhou, J. Brassinosteroid ameliorates zinc oxide nanoparticlesinduced oxidative stress by improving antioxidant potential and redox homeostasis in tomato seedling. *Front. Plant Sci.* 2016, 7, 1–13. [CrossRef]
- Dimkpa, C.O.; Bindraban, P.S. Nanofertilizers: New products for the industry? J. Agric. Food Chem. 2018, 66, 6462–6473. [CrossRef] [PubMed]

- Nemec, S.; Kralj, S.; Wilhelm, C.; Abou-Hassan, A.; Rols, M.P.; Kolosnjaj-Tabi, J. Comparison of iron oxide nanoparticles in photothermia and magnetic hyperthermia: Effects of clustering and silica encapsulation on nanoparticles' heating yield. *Appl. Sci.* 2020, 10, 7322. [CrossRef]
- Emamverdian, A.; Ding, Y.; Mokhberdoran, F.; Ahmad, Z.; Xie, Y. The Investigation of TiO₂NPs Effect as a Wastewater Treatmentto Mitigate Cd Negative Impact on Bamboo Growth. *Sustainability* 2021, *13*, 3200. [CrossRef]
- 14. Emamverdian, A.; Ding, Y.; Mokhberdoran, F.; Ramakrishnan, M.; Ahmad, Z.; Xie, Y. Different Physiological and Biochemical Responses of Bamboo to the Addition of TiO₂ NPs under Heavy Metal Toxicity. *Forests* **2021**, *12*, 759. [CrossRef]
- Rizwan, M.; Ali, S.; ur Rehman, M.Z.; Malik, S.; Adrees, M.; Qayyum, M.F.; Alamri, S.A.; Alyemeni, M.N.; Ahmad, P. Effect of foliar applications of silicon and titanium dioxide nanoparticles on growth, oxidative stress, and cadmium accumulation by rice (*Oryza sativa*). Acta Physiol. Plant 2019, 41, 35. [CrossRef]
- Rehana Sardar, G.; Ahmed, S.; Ahmad Yasin, N. Titanium dioxide nanoparticles mitigate cadmium toxicity in *Coriandrum sativum* L. through modulating antioxidant system, stress markers and reducing cadmium uptake. *Environ. Pollut* 2022, 292, 118373. [CrossRef]
- 17. Faraji, J.; Sepehri, A. Titanium Dioxide Nanoparticles and Sodium Nitroprusside Alleviate the Adverse Effects of Cadmium Stress on Germination and Seedling Growth of Wheat (*Triticum aestivum* L.). *Univ. Sci.* **2018**, 23, 61–87. [CrossRef]
- 18. Emamverdian, A.; Ding, Y.; Xie, Y. The Role of New Members of Phytohormones in Plant Amelioration under Abiotic Stress with an Emphasis on Heavy Metals. *Pol. J. Environ. Stud.* **2020**, *29*, 1009–1020. [CrossRef]
- 19. Zafari, M.; Ebadi, A.; Sedghi, M.; Jahanbakhsh, S.; Miransari, M.J. Alleviating effect of 24-epibrassinolide on seed oil content and fatty acid composition under drought stress in safflower. *J. Food Compost. Anal.* **2020**, *92*, 103544. [CrossRef]
- Siddiqui, H.; Hayat, S.; Bajguz, A. Regulation of photosynthesis by brassinosteroids in plants. *Acta Physiol. Plant.* 2018, 40, 11738–12018. [CrossRef]
- Hayat, S.; Hasan, S.A.; Yusuf, M.; Hayat, Q.; Ahmad, A. Effect of 28-homobrassinolide on photosynthesis, fluorescence and antioxidant system in the presence or absence of salinity and temperature in *Vigna radiata*. *Environ. Exp. Bot.* 2010, 69, 105–112. [CrossRef]
- 22. Ahanger, M.A.; Ashraf, M.; Bajguz, A.; Ahmad, P. Brassinosteroids regulate growth in plants under stressful environments and crosstalk with other potential phytohormones. *J. Plant Growth Regul.* **2018**, *37*, 1007–1024. [CrossRef]
- Zhou, J.; Liu, D.; Wang, P.; Ma, X.; Lin, W.; Chen, S.; Mishev, K.; Lu, D.; Kumar, R.; Vanhoutte, I.; et al. Regulation of Arabidopsis brassinosteroid receptor BRI1 endocytosis and degradation by plant U-box PUB12/PUB13-mediaed ubiquitination. *Proc. Natl. Acad. Sci. USA* 2018, 115, 1906–1915. [CrossRef]
- 24. Li, Y.M.; Feng, P.F. Bamboo resources in china based on the ninth national forest inventory data. *World Bamboo Ratt.* **2019**, *17*, 45–48.
- 25. Kang, F.; Li, X.; Du, H.; Mao, F.; Zhou, G.; Xu, Y.; Huang, Z.; Ji, J.; Wang, J. Spatiotemporal Evolution of the Carbon Fluxes from Bamboo Forests and their Response to Climate Change Based on a BEPS Model in China. *Remote Sens.* **2022**, *14*, 366. [CrossRef]
- Emamverdian, A.; Ding, Y.; Ranaei, F.; Ahmad, Z. Application of bamboo plants in nine aspects. Sci. World J. 2020, 2020, 7284203. [CrossRef]
- Huang, W.; Olson, E.; Wang, S.H.; Shi, P. The growth and mortality of *Pleioblastus pygmaeus* under different light availability. *Glob. Ecol.* 2020, 24, e01262. [CrossRef]
- Murashige, T.; Skoog, F. A Revised Medium for Rapid Growth and Bio Assays with Tobacco Tissue Cultures. *Physiol. Plant.* 1962, 15, 473–497. [CrossRef]
- Zhang, X.Z. The Measurement and Mechanism of Lipid Peroxidation and SOD, POD and CAT Activities in Biological System. In Research Methodology of Crop Physiology; Agriculture Press: Beijing, China, 1992; pp. 208–211.
- Upadhyaya, A.; Sankhla, D.; Davis, T.D.; Sankhla, N.; Smith, B.N. Effect of paclobutrazol on the activities of some enzymes of activated oxygen metabolism and lipid peroxidation in senescing soybean leaves. J. Plant Physiol. 1985, 121, 453–461. [CrossRef]
- Aebi, H. Catalase in vitro. In *Methods in Enzymology*; Elsevier: Amsterdam, The Netherlands, 1984; Volume 105, pp. 121–126. [CrossRef]
- 32. Foyer, C.H.; Halliwell, B. The presence of glutathione and glutathione reductase in chloroplasts: A proposed role in ascorbic acid metabolism. *Planta* **1976**, *133*, 1–25. [CrossRef]
- Nakano, Y.; Asada, K. Hydrogen peroxide is scavenged by ascorbate-specific peroxidase in spinach chloroplasts. *Plant Cell Physiol.* 1981, 22, 867–880. [CrossRef]
- 34. Berner, M.; Krug, D.; Bihlmaier, C.; Vente, A.; Muller, R.; Bechthold, A. Genes and enzymes involved in caffeic acid biosynthesis in Actinomycete *Saccharothrix espanaensis*. *J. Bacteriol.* **2006**, *188*, 2666–2673. [CrossRef]
- 35. Conde, E.; Cadahia, E.; Garcia-Vallejo, M. HPLC analysis of flavonoids and phenolic acids and aldehydes in *Eucalyptus* spp. *Chromatographia* **1995**, *41*, 657–660. [CrossRef]
- Akkol, E.K.; Goger, F.; Koşar, M.; Başer, K.H.C. Phenolic composition and biological activities of Salvia halophila and Salvia virgata from Turkey. *Food Chem.* 2008, 108, 942–949. [CrossRef] [PubMed]
- Kayden, H.J.; Chow, C.K.; Bjornson, L.K. Spectrophotometric method for determination of tocopherol in red blood cells. J. Lipid Res. 1973, 14, 533–540. [CrossRef]

- Siddiqui, H.; Ahmad, K.B.M.; Hayat, S. Comparative effect of 28-homobrassinolide and 24-epibrassinolide on the performance of different components influencing the photosynthetic machinery in *Brassica juncea* L. *Plant Physiol. Biochem* 2018, 129, 198–212. [CrossRef] [PubMed]
- Patterson, B.D.; MacRae, E.A.; Ferguson, I.B. Estimation of hydrogen peroxide in plant extracts using titanium (IV). *Anal. Biochem.* 1984, 139, 487–492. [CrossRef]
- Bradford, M.M. A rapid sensitive method for the quantification of microgram quantities of protein utilising the principle of protein-Dye Binding. *Anal. Biochem.* 1976, 72, 248–254. [CrossRef]
- 41. Li, C.; Bai, T.; Ma, F.; Han, M. Hypoxia tolerance and adaptation of anaerobic respiration to hypoxia stress in two Malus species. *Sci. Hortic.* **2010**, 124, 274–279. [CrossRef]
- 42. Valentovic, P.; Luxova, M.; Kolarovic, L.; Gasparikova, O. Effect of osmotic stress on compatible solutes content, membrane stability and water relations in two maize cultivars. *Plant Soil. Environ.* **2006**, *52*, 186–191. [CrossRef]
- 43. Lichtenthaler, H.K.; Buschmann, C. Chlorophylls and carotenoids: Measurement and characterization by UV–VIS specroscopy. In *Current Protocols in Food Analytical Chemistry*; John Wiley & Sons, Inc.: Hoboken, NJ, USA, 2001; p. 4.
- 44. Souri, Z.; Karimi, N. Enhanced phytoextraction by as hyperaccumulator *Isatis cappadocica* spiked with sodium nitroprusside. *Soil Sediment Contam. Int. J.* 2017, 26, 457–468. [CrossRef]
- 45. Wang, J.; Nie, Y.; Dai, H.; Wang, M.; Cheng, L.; Yang, Z.; Chen, S.; Zhao, G.; Wu, L.; Guang, S. Parental exposure to TiO₂ NPs promotes the multigenerational reproductive toxicity of Cd in Caenorhabditis elegans via bioaccumulation of Cd in germ cells. *Environ. Sci. Nano* 2019, *6*, 1332–1342. [CrossRef]
- Singh, D.; Kumar, S.; Singh, S.C.; Lal, B.; Singh, N.B. Applications of liquid assisted pulsed laser ablation synthesized TiO₂ nanoparticles on germination, growth and biochemical parameters of *Brassica oleracea* var. *Capitata. Sci. Adv. Mather.* 2012, 4, 522–531. [CrossRef]
- Bajguz, A.; Hayat, S. Effects of brassinosteroids on the plant responses to environmental stresses. *Plant Physiol. Biochem.* 2009, 47, 1–8. [CrossRef] [PubMed]
- Bajguz, A. Brassinosteroids and lead as stimulators of phytochelatins synthesis in *Chlorella vulgaris*. J. Plant Physiol. 2002, 159, 321–324. [CrossRef]
- Sharma, I.; Pati, P.K.; Bhardwaj, R. Effect of 28-homobrassinolide on antioxidant defence system in *Raphanus sativus* L. under chromium toxicity. *Ecotoxicology* 2011, 20, 862–874. [CrossRef]
- Arora, P.; Bhardwaj, R.; Kanwar, M.K. Effect of 24-epibrassinolide on growth, protein content and antioxidative defense system of Brassica juncea L. subjected to cobalt ion toxicity. Acta Physiol. Plant 2012, 34, 2007–2017. [CrossRef]
- 51. Kohli, S.K.; Handa, N.; Sharma, A.; Gautam, V.; Arora, S.; Bhardwaj, R.; Alyemeni, M.N.; Wijaya, L.; Ahmad, P. Combined effect of 24-epibrassinolide and salicylic acid mitigates lead (Pb) toxicity by modulating various metabolites in *Brassica juncea* L. seedlings. *Protoplasma* **2018**, *255*, 11–24. [CrossRef]
- Ahmad, P.; Abdel Latef, A.A.; Abd_Allah, E.F.; Hashem, A.; Sarwat, M.; Anjum, N.A.; Gucel, S. Calcium and potassium supplementation enhanced growth, osmolyte secondary metabolite production, and enzymatic antioxidant machinery in cadmium-exposed chickpea (*Cicer arietinum* L.). *Front. Plant Sci.* 2016, 7, 513. [CrossRef]
- 53. Choudhary, S.P.; Kanwar, M.; Bhardwaj, R.; Yu, J.-Q.; Tran, L.-S.P. Chromium stress mitigation by polyamine-brassinosteroid application involves phytohormonal and physiological strategies in *Raphanus sativus* L. *PLoS ONE* **2012**, *7*, e33210. [CrossRef]
- 54. Deng, X.-G.; Zhu, T.; Peng, X.-J.; Xi, D.-H.; Guo, H.; Yin, Y.; Zhang, D.-W.; Lin, H.-H. Role of brassinosteroid signaling in modulating tobacco mosaic virus resistance in *Nicotiana benthamiana*. *Sci. Rep.* **2016**, *6*, 20579. [CrossRef] [PubMed]
- 55. Mierziak, J.; Kostyn, K.; Kulma, A. Flavonoids as important molecules of plant interactions with the environment. *Molecules* **2014**, 19, 16240–16265. [CrossRef] [PubMed]
- Kováčik, J.; Bačkor, M. Phenylalanine ammonia-lyase and phenolic compounds in chamomile tolerance to cadmium and copper excess. Water Air Soil Pollut. 2007, 185, 185–193. [CrossRef]
- 57. Asghari, M.; Zahedipour, P. 24-Epibrassinolide acts as a growth-promoting and resistance-mediating factor in strawberry plants. *J. Plant Growth Regul.* **2016**, *35*, 722–729. [CrossRef]
- Manquián-Cerda, K.; Escudey, M.; Zúñiga, G.; Arancibia-Miranda, N.; Molina, M.; Cruces, E. Effect of cadmium on phenolic compounds, antioxidant enzyme activity and oxidative stress in blueberry (*Vaccinium corymbosum* L. plantlets grown in vitro. *Ecotoxicol. Environ. Saf.* 2016, 133, 316–326. [CrossRef] [PubMed]
- 59. Sanjari, S.; Keramat, B.; Nadernejad, N.; Mozafari, H. Ameliorative effects of 24-epibrassinolide and thiamine on excess cadmiuminduced oxidative stress in canola (*Brassica napus* L.) plants. J. Plant Interact. 2019, 14, 359–368. [CrossRef]
- 60. Mohammadi, R.; MaaliAmiri, R.; Mantri, N.L. Effect of TiO₂ nanoparticles on oxidative damage and antioxidant defense systems in chickpea seedlings during cold stress. *Russ. J. Plant Physiol.* **2014**, *61*, 768–775. [CrossRef]
- Demin, I.N.; Deryabin, A.N.; Sinkevich, M.S.; Trunova, T.I. Insertion of cyanobacterial *desA* gene coding for Δ12-acyl-lipid desaturase increases potato plant resistance to oxidative stress induced by hypothermia. *Russ. J. Plant Physiol.* 2008, 55, 639–648. [CrossRef]
- 62. Surgun, Y.; Çöl, B.; Bürün, B. 24-Epibrassinolide ameliorates the effects of boron toxicity on *Arabidopsis thaliana* (L.) Heynh by activating an antioxidant system and decreasing boron accumulation. *Acta Physiol. Plant* **2016**, *38*, 71. [CrossRef]
- 63. Talarek-Karwel, M.; Bajguz, A.; Piotrowska-Niczyporuk, A. 24-Epibrassinolide modulates primary metabolites, antioxidants, and phytochelatins in *Acutodesmus obliquus* exposed to lead stress. *J. Appl. Phycol.* **2020**, *32*, 263–276. [CrossRef]

- 64. Santos, L.R.; Batista, B.L.; Lobato, A.K.S. Brassinosteroids mitigate cadmium toxicity in cowpea plants. *Photosynthetica* 2018, *56*, 591–605. [CrossRef]
- 65. Pereira, Y.C.; Rodrigues, W.S.; Lima, E.J.A.; Santos, L.R.; Silva, M.H.L.; Lobato, A.K.S. Brassinosteroids increase electron transport and photosynthesis in soybean plants under water deficit. *Photosynthetica* **2019**, *57*, 181–191. [CrossRef]
- 66. Fariduddin, Q.; Yusuf, M.; Ahmad, I.; Ahmad, A. Brassinosteroids and their role in response of plants to abiotic stresses. *Biol. Plant.* **2014**, *58*, 9–17. [CrossRef]
- 67. Lei, Z.; Minqyu, S.; Xiao, W.; Chao, L.; Chunxiang, Q.; Liang, C.; Hao, H.; Xiaoqing, L.; Fashui, H. Antioxidant stress is promoted by nano-anatase in spinach chloroplasts under UVB radiation. *Biol. Trace Elem. Res.* **2008**, *121*, 69–79. [CrossRef] [PubMed]
- 68. Talarek-Karwel, M.; Bajguz, A.; Piotrowska-Niczyporuk, A.; Rajewska, I. The effect of 24-epibrassinolide on the green alga *Acutodesmus obliquus* (Chlorophyceae). *Plant Physiol. Biochem.* **2018**, *124*, 175–183. [CrossRef] [PubMed]
- 69. Bajguz, A. Effect of brassinosteroids on nucleic acids and protein content in cultured cells of *Chlorella vulgaris*. *Plant Physiol*. *Biochem*. **2000**, *38*, 209–215. [CrossRef]
- Jan, S.; Alyemeni, M.N.; Wijaya, L.; Alam, P.; Siddique, K.H.; Ahmad, P. Interactive effect of 24-epibrassinolide and silicon alleviates cadmium stress via the modulation of antioxidant defense and glyoxalase systems and macronutrient content in *Pisum sativum* L. seedlings. *BMC Plant Biol.* 2018, 18, 146. [CrossRef]
- Lyu, S.; Wei, X.; Chen, J.; Wang, C.; Wang, X.; Pa, D. Titanium as a beneficial element for crop production. *Front. Plant Sci.* 2017, *8*, 597. [CrossRef]
- 72. Bacilieri, F.S.; Pereira de Vasconcelos, A.C.; Quintao Lana, R.M.; Mageste, J.G.; Torres, J.L.R. Titanium (Ti) in plant nutrition—A review. *Aust. J. Crop Sci.* 2017, *11*, 382–386. [CrossRef]
- 73. Daryabeigi Zand, A.; Mikaeili Tabrizi, A.; Vaezi Heir, A. Co-application of biochar and titanium dioxide nanoparticles to promote remediation of antimony from soil by Sorghum bicolor: Metal uptake and plant response. *Heliyon* **2020**, *6*, e04669. [CrossRef]
- 74. Bajguz, A. Suppression of Chlorella vulgaris growth by cadmium, lead, and copper stress and Its restoration by endogenous brassinolide. *Arch. Environ. Contam. Toxicol.* **2011**, *60*, 406–416. [CrossRef] [PubMed]
- 75. Kanwar, M.K.; Bhardwaj, R.; Arora, P.; Chowdhary, S.P.; Sharma, P.; Kumar, S. Plant steroid hormones produced under Ni stress are involved in the regulation of metal uptake and oxidative stress in *Brassica juncea* L. *Chemosphere* 2012, *86*, 41–49. [CrossRef] [PubMed]
- 76. Kroutil, M.; Hejtmánková, A.; Lachman, J. Effect of spring wheat (*Triticum aestivum* L.) treatment with brassinosteroids on the content of cadmium and lead in plant aerial biomass and grain. *Plant Soil Environ.* 2010, *56*, 43–50. [CrossRef]
- 77. Dong, Y.; Chen, W.; Bai, X.; Liu, F.; Wan, Y. Effects of Exogenous Nitric Oxide and 24-Epibrassinolide on the Physiological Characteristics of Peanut Seedlings Under Cadmium Stress. *Pedosphere* **2019**, *29*, 45–59. [CrossRef]
- 78. Wani, A.S.; Tahir, I.; Ahmad, S.S.; Dar, R.A.; Nisar, S. Efficacy of 24-epibrassinolide in improving the nitrogen metabolism and antioxidant system in chickpea cultivars under cadmium and/or NaCl stress. *Sci. Hortic.* **2017**, 225, 48–55. [CrossRef]
- Ahmad, P.; Ahanger, M.A.; Alyemeni, M.N.; Wijaya, L.; Alam, P. Exogenous application of nitric oxide modulates osmolyte metabolism, antioxidants, enzymes of ascorbate-glutathione cycle and promotes growth under cadmium stress in tomato. *Protoplasma* 2018, 255, 79–93. [CrossRef]
- 80. Kelemen, G.; Keresztes, A.; Bacsy, E.; Feher, M.; Fodor, P.; Pais, I. Distribution and intracellular localization of titanium in plants after titanium treatment. *Food Struct.* **1993**, *12*, 8.
- Silva, S.; Craveiro, S.C.H.; Oliveira, A.J.; Calado, R.J.B.; Pinto, A.M.S.; Silva, C. Santos Wheat chronic exposure to TiO₂nanoparticles: Cyto- and genotoxic approach. *Plant Physiol. Biochem.* 2017, 121, 89–98. [CrossRef]
- Larue, C.; Laurette, J.; Herlin-Boime, N.; Khodja, H.; Fayard, B.; Flank, A.M.; Brisset, F.M. Carriere Accumulation, translocation and impact of TiO₂ nanoparticles in wheat (*Triticum aestivum* spp.): Influence of diameter and crystal phase. *Sci. Total Environ.* 2012, 431, 197–208. [CrossRef]
- 83. Rahneshan, Z.; Nasibi, F.; Moghadam, A.A. Effects of salinity stress on some growth, physiological, biochemical parameters and nutrients in two pistachio (*Pistacia vera* L.) rootstocks. *J. Plant. Interact.* **2018**, *13*, 73–82. [CrossRef]
- Bajguz, A. An enhancing effect of exogenous brassinolide on the growth and antioxidant activity in Chlorella vulgaris cultures under heavy metals stress. *Environ. Exp. Bot.* 2010, 68, 175–179. [CrossRef]
- Rasool, S.; Urwat, U.; Nazir, M.; Zargar, S.M.; Zargar, M.Y. Cross talk between phytohormone signaling pathways under abiotic stress conditions and their metabolic engineering for conferring abiotic stress tolerance. In *Abiotic Stress-Mediated Sensing and Signaling in Plants: An Omics Perspective*; Zargar, S.M., Zargar, M.Y., Eds.; Springer: New York, NY, USA, 2018; pp. 329–350.
- Tripathi, B.N.; Mehta, S.K.; Amar, A.; Gaur, J.P. Oxidative stress in Scenedesmus sp. during short- and long-term exposure to Cu²⁺ and Zn²⁺. *Chemosphere* 2006, *62*, 538–544. [CrossRef] [PubMed]
- Polonini, H.C.; Brandao, H.M.; Raposo, N.R.; Brandao, M.A.; Mouton, L.; Coute, A.; Yepremian, C.; Sivry, Y.; Brayner, R. Size-dependent ecotoxicity of barium titanate particles: The case of *Chlorella vulgaris* green algae. *Ecotoxicology* 2015, 24, 938–948. [CrossRef] [PubMed]
- Guo, J.; Zhou, R.; Ren, X.; Jia, H.; Hua, L.; Xu, H.; Lv, X.; Zhao, J.; Wei, T. Effects of salicylic acid, epi-brassinolide and calcium on stress alleviation and Cd accumulation in tomato plants. *Ecotoxicol. Environ. Saf.* 2018, 157, 491–496. [CrossRef] [PubMed]
- Yusuf, M.; Khan, T.A.; Fariduddin, Q. Interaction of epibrassinolide and selenium ameliorates the excess copper in *Brassica juncea* through altered proline metabolism and antioxidants. *Ecotoxicol. Environ. Saf.* 2016, 129, 25–34. [CrossRef]

- 90. Wu, C.; Li, F.; Xu, H.; Zeng, W.; Yu, R.; Wu, X.; Shen, L.; Liu, Y.; Li, J. The potential role of brassinosteroids (BRs) in alleviating antimony (Sb) stress in *Arabidopsis thaliana*. *Plant Physiol. Biochem.* **2019**, *141*, 51–59. [CrossRef]
- 91. Lima, M.D.R.; Barros, U.D.; Batista, B.L.; Lobato, A.K.S. Brassinosteroids mitigate iron deficiency improving nutritional status and photochemical efficiency in *Eucalyptus urophylla* plants. *Trees Struct. Funct.* **2018**, *32*, 1681–1694. [CrossRef]
- Li, Y.; Song, Y.; Shi, G.; Wang, J.; Hou, X. Response of antioxidant activity to excess copper in two cultivars of Brassica campestris ssp. chinensis Makino. Acta Physiol. Plant. 2008, 31, 155–162. [CrossRef]
- 93. Saeidnejad, A.; Mardani, H.; Naghibolghora, M. Protective effects of salicylic acid on physiological parameters and antioxidants response in maize seedlings under salinity stress. J. Appl. Environ. Biol. Sci. 2012, 2, 364–373.
- Singh, J.; Lee, B.K. Influence of nano-TiO₂ particles on the bioaccumulation of Cd in soybean plants (*Glycine max*): A possible mechanism for the removal of Cd from the contaminated soil. *J. Environ. Manag.* 2016, 170, 88–96. [CrossRef]
- Abdel Latef, A.A.H.; Srivastava, A.K.; El-sadek, M.S.A.; Kordrostami, M.; Tran, L.S.P. Titanium Dioxide Nanoparticles Improve Growth and Enhance Tolerance of Broad Bean Plants under Saline Soil Conditions. *Land Degrad. Dev.* 2017, 29, 1065–1073. [CrossRef]
- 96. Li, F.M.; Zhao, W.; Li, Y.Y.; Tian, Z.J.; Wang, Z.Y. Toxic effects of nanoTiO₂ on *Gymnodinium breve*. Environ. Sci. 2012, 33, 233–238.
- Feizi, H.; Moghaddam, P.R.; Shahtahmassebi, N.; Fotovat, A. Impact of bulk and nanosized titanium dioxide (TiO₂) on wheat seed germination and seedling growth. *Biol. Trace. Elem. Res.* 2012, 146, 101–106. [CrossRef] [PubMed]
- 98. Tan, W.; Peralta-Videa, J.R.; Gardea-Torresdey, J.L. Interaction of titanium dioxide nanoparticles with soil components and plants: Current knowledge and future research needs—A critical review. *Environ. Sci. Nano* **2018**, *5*, 257–278. [CrossRef]
- Mattiello, A.; Filippi, A.; Pošćić, F.; Musetti, R.; Salvatici, M.C.; Giordano, C.; Vischi, M.; Bertolini, A.; Marchiol, L. Evidence of phytotoxicity and genotoxicity in *Hordeum vulgare* L. exposed to CeO₂ and TiO₂ nanoparticles. *Front. Plant. Sci.* 2015, 6, 1043. [CrossRef]
- Rastogi, A.; Zivcak, M.; Sytar, O.; Kalaji, H.M.; He, X.; Mbarki, S.; Brestic, M. Impact of Metal and Metal Oxide Nanoparticles on Plant: A Critical Review. Front. Chem. 2017, 5, 78. [CrossRef]
- Kobayashi, K.; Kubota, H.; Hojo, R.; Miyagawa, M. Effective dispersal of titanium dioxide nanoparticles for toxicity testing. *J. Toxicol. Sci.* 2019, 44, 515–521. [CrossRef]